

Muncie Sanitary district

Indiana Scientific Purpose License Number: 18-161

Bureau of Water Quality Annual Macroinvertebrate Community Report 2018

Bureau of Water Quality 5150 W. Kilgore Ave. Muncie, IN 47304

Phone: 765-747-4896 Fax: 765-213-6444 www.munciesanitary.org/bwq

Prepared by: Laura Bowley, Macroinvertebrate Biologist, BWQ March 2019

Photo description (previous page): Lampsilis fasciola showing one of its four known lure displays. All four displays were seen at a single site in 2018.

TABLE OF CONTENTS	. 3
PREFACE	. 5
INTRODUCTION	. 5
West Fork White River and the Bureau of Water Quality	. 5
Mussels as Biomonitors	. 6
Figure 1.— Mussel sampling segments, 2018.	. 6
Macroinvertebrates as Biomonitors	. 7
Figure 2.— Macroinvertebrates sites, 2018	. 7
MUSSEL METHODS	. 8
Mussels- Field Sampling	. 8
Table 1.—mIBI submetrics and their response to disturbance	. 8
MACROINVERTEBRATE METHODS	. 8
Macroinvertebrate Field Sampling	. 8
Table 2.—mIBI scores and corresponding ratings	. 9
Macroinvertebrate Laboratory Methods	. 9
Macroinvertebrate Data Tabulation	. 9
IDEM's Macroinvertebrate Index of Biotic Integrity (mIBI)	. 9
Hilsenhoff Biotic Index (HBI)	. 9
Table 3.—HBI values and corresponding ratings	. 9
Shannon-Wiener Diversity Index (H')	. 9
Table 4.—QHEI scores and corresponding ratings.	10
Shannon Evenness Index (J')	10
Percent Dominance of Top Three Taxa	10
Percent Chironomidae	10
Qualitative Habitat Evaluation Index (QHEI)	10
MUSSEL RESULTS	10
MACROINVERTEBRATE RESULTS	10
mIBI.—White River	10
mIBI.—Buck Creek	10
Graph 1.—White River mIBI scores, 2018	11
Graph 2.—Buck Creek mIBI scores, 2018	11
Graph 3.—Smaller tributary mIBI scores, 2018	11
mIBI.—Smaller Tributaries	11
Stand Alone Indices	11
HBI: White River	11
HBI: Buck Creek	11
HBI: Smaller Tributaries	11
H': White River	11
H': Buck Creek	11
H': Smaller Tributaries	11
Graph 4.—White River HBI scores, 2018	12
Graph 5.—Buck Creek HBI scores, 2018	12

TABLE OF CONTENTS

Graph 6.—Smaller tributary HBI scores, 2018	12
Remaining Stand Alone Indices: White River	12
Remaining Stand Alone Indices: Buck Creek	12
Remaining Stand Alone Indices: Smaller Tributaries	12
Graph 7.—White River H' scores, 2018	
Graph 8.—Buck Creek H' scores, 2018	13
Graph 9—Smaller tributary H' scores, 2018	13
QHEI: White River	13
QHEI: Buck Creek	13
QHEI: Smaller Tributaries	13
DISCUSSION	13
Mussels	13
Graph 10.—White River QHEI scores, 2018	14
Graph 11.—Buck Creek QHEI scores, 2018	14
Graph 13.—Smaller tributary QHEI scores, 2018	14
Macroinvertebrates	14
Appendix A.— Mussels	17
Table 4.—Mussel assemblages within Muncie city limits, 2018	18
Graph 13.—Relative abundance for mussels within Muncie city limits, 2018	18
Appendix B.— Macroinvertebrates	19
Table 5.—Macroinvertebrate field sheets	20
Table 6.—Macroinvertebrate site descriptions and locations, 2018	22
Table 7.—Tolerance values and attributes used in mIBI and HBI calculations	26
Table 8.—Macroinvertebrate site scores, 2018	31
Table 9.—Mean scores for macroinvertebrate metrics, 2018	
Table 10.—Macroinvertebrate field sheets	
REFERENCES	35

Acknowledgements: Thank you to Emily Musenbrock and Kevin Meyer for their assistance in obtaining these samples, and for their countless hours of data entry, proofing, slide preparation, and general assistance in preparing this report. Extra thanks and acknowledgement to Kevin Meyer for creating Figure 1.

PREFACE

This paper contains results of the Bureau of Water Quality's (BWQ's) macroinvertebrate and mussel biomonitoring for the year 2018. For the purpose of displaying trends, some graphs and tables will present data from past years. However, the analysis given here is only for 2018. If further investigation of past years is needed, please refer to prior reports from this organization.

From 2013-2018 an additional Buck Creek site was sampled. This site (BUC 0.0) was sampled to observe changes in the site before and after best management practices (implemented in late 2013) were put into place.

In 2014, one zebra mussel *Dreissena polymorpha* was found on a sampler in Prairie Creek Reservoir, upstream of Muncie. The reservoir is very near White River, connected via Prairie Creek. In 2015 zebra mussels were found on a sampler in Prairie Creek. In 2018 zebra mussels are well established throughout the West Fork White River downstream of Prairie Creek.

Unique to this year, no "sites" were sampled for mussels. Instead, a project was undertaken to complete timed surveys on surface mussels in the West Fork White River throughout Delaware County. In 2018, this was completed within Muncie city limits. This will likely be a three to four year project.

INTRODUCTION

West Fork White River and the Bureau of Water Quality.—The headwaters of the West Fork White River (WFWR) can be found near Winchester, Indiana, moving westward through Muncie, draining approximately 384 square miles at the Madison County/Delaware County line (Hoggat 1975). The land along the river in Delaware County is primarily used for agriculture (corn, soybeans, and livestock), but also includes the urban area of Muncie. Muncie is a heavily industrialized community that has included electroplating firms, transmission assembly plants, a secondary lead smelter, foundries, heat treatment operations, galvanizing operations, and tool and die shops (ICLEI Case Study #19 1994).

In 1972, the Division of Water Quality (DWQ), now named the Bureau of Water Quality (BWQ), was established out of a need to regulate and control the sources responsible for polluting White River and its tributaries in and around Muncie, Indiana. The BWQ also wanted to attain those goals set forth by legislation of the 1970's and 1980's (The Water Pollution Act of 1972, the Clean Water Act of 1977 and the Water Quality Act of 1987). One of the ultimate goals is biological integrity, defined by Karr & Dudley (1981) as "the ability to support and maintain a balanced, integrated, adaptive community of organisms having a species composition, diversity, and functional organization comparable to that of natural habitat of the region."

Since the establishment of the BWQ, industries have installed millions of dollars in industrial pretreatment equipment, and corrective action is constantly being taken to prevent spills from entering the sewers and waterways. In addition, an ongoing program has reduced, and in some cases eliminated, pollution entering White River from combined sewer overflows (CSOs). Improvements have been made to the Muncie Water Pollution Control Facility (MWPCF), local sewers have been built to correct septic tank problems, and wildlife habitat has been developed along the river (Craddock 1990).

To get the best representation of the quality of a water system, both chemical and biological

monitoring should be implemented. The benefits of chemical testing are vast; however, chemical monitoring can miss or underestimate combined chemical effects, sporadic events, and other factors such as habitat degradation (Karr 1981).

A benefit to using biological communities as indicators of water quality is their longevity and sensitivity to disturbances in the habitat in which they live. The observed condition of the aquatic biota, at any given time, is the result of the chemical and physical dynamics that occur in a water body over time (OEPA DWQMA 1987). Alone, neither gives a complete picture of water quality, however, the combination of biological and chemical monitoring increases the chances that degradation to the water body will be detected (Karr 1991).

Mussels as biomonitors.—Freshwater mussels are considered the most imperiled group of organisms in North America (Lydeard et al. 2004; Strayer et al. 2004), if not the world (Strayer 2008), and are declining at alarming and unprecedented rates (Neves et al 1997; Ricciardi & Rasmussen 1999; Vaughn & Taylor 1999; Strayer & Smith 2003; Poole & Downing 2004; Regnier et al. 2009). In North America alone, 72% of the native mussel fauna is either federally listed as endangered or threatened or considered to be in need of some protection (Haag 2009). At one time, 90 species of Unionid (of the family Unionidae) mussels were known to have existed in

Figure 1.—Mussel sampling sections, 2018.

the eight Great Lake and Upper Mississippi states. Now, 33% are listed as extinct, endangered, or are candidates for that listing (Ball & Schoenung 1995). In the United States, 71 taxa are currently listed as endangered or threatened by the Endangered Species Act (USFWS 2005) and are suffering an extinction rate higher than any other North American fauna (Ricciardi & Rasmussen 1999). Contributors to this decline include commercial harvest, degradation of habitat (including channelization and dredging), toxic and siltation. Other significant chemicals, contributors include: impoundments (Vaughn & Taylor 1999; Watters 2000; Dean et al. 2002), water pollution (organic, inorganic, and thermal) (Mummert et al. 2003; Keller & Augspurger 2005; Valenti et al. 2005; 2006; Gooding et al. 2006; Bringolf et al. 2007; March et al. 2007; Wang et al. 2007; Cope et al. 2008; Besser et al. 2009), habitat alterations, and land use practices (Clarke 1981; Ball & Schoenung 1995; Biggins et al. 1995; Couch 1997; Gatenby et al. 1998; Payne et al. 1999; Watters 1999; Poole & Downing 2004). In 1990, the US EPA listed sedimentation as the top pollutant of rivers in the United States (Box & Mossa 1999). Studies have shown that silt accumulation of 0.25 to 1 inch resulted in nearly 90% mortality of mussels tested (Ellis 1936). This affects mussels by reducing interstitial flow rates, clogging mussel gills, and reducing light for photosynthesis of algae (primary forage of the



mussel). Suspended particles also cause difficulty with the necessary fish and mussel interactions needed for reproduction and survival (Box & Mossa 1999). These indicate the importance of water quality as a factor in mussel survival. It is for these reasons, as well as their long life span, feeding habits, persistent shells (Strayer 1999a) and sensitive growth and reproductive rates (Burky 1983) that mussels serve well as biological indicators.

Macroinvertebrates as Biomonitors.—There are numerous reasons for using macroinvertebrates as indicators of water quality. Their ubiquitous nature, large numbers (individuals and species), and relative ease of sampling with inexpensive equipment make them ideal for bioassessments (Lenat et al. 1980; Hellawell 1986; Lenat & Barbour 1993). Macroinvertebrates are relatively sessile, allowing spatial analysis of disturbances (Tesmer & Wefring 1979; Hellawell 1986; Abel 1989). The extended life cycles of most aquatic insects allows for temporal analysis as well (Lenat et al. 1980; Hellawell 1986). Finally, macroinvertebrate species are well documented; many identification keys and forms of analysis are available, and specific responses to pollutants and stressors are well known (Hellawell 1986; Abel 1989; Rosenberg & Resh 1993). They are especially useful in situations where intermittent or mild

Macro Sites 2018 2018 Albany mIBI Excellent Good Fair Poor Very Poor 2018 HBI Excellent arkeParker City Very Good Good Fair Fairly Poor Poor 2018 QHEI Palev Excellent Good Fair Middletowr Bountsvill Poor SpringBRA Very Poor Sources: Esri, HERE, DeLorme, Intermap, incrementer Corp., GEBCO, USGS, FAO, NPS, NRCAN, GeoBase, IGN, Kadaster NL, Ordnance Survey Esri Japan, METI, Esri China (Hong Kong), swisstoppur MapmyIndia, © OpenStreetMap contributors and the GIS User Community. Esri, HERE, DeLorme, MapmyIndia, © OpenStreetMap contributors and the GIS user community 0 1.5 6 Miles 3 Muncie City Limits

Figure 2.—Macroinvertebrate sites, 2018.

organic enrichment is present (Chutter 1972).

MUSSEL METHODS

Mussel Field Sampling.— In 2018, a project was begun to determine the distribution of mussels on White River from the Delaware/Randolph County line to the Delaware/Madison county line. Sampling methods followed the Timed Search Survey. This is one of the more popular sampling methods, due to its efficient coverage of large areas (Metcalf-Smith et al 2000), and its effectiveness at obtaining high species richness and finding rare species (Vaughn et al 1997).

Sampling began at the downstream city limits of Muncie, and proceeded upstream. Densities were determined using catch per unit effort.

MACROINVERTEBRATE METHODS

Macroinvertebrate Field Sampling.— Macroinvertebrate samples were taken at 14 sites on White River, and five sites along Buck Creek (Figure 1 and Appendix B, Table 9). Sampling followed the current IDEM Multi-habitat Macroinvertebrate Collection Procedure (MHAB) (IDEM 2010). This methodology includes a composite of a one minute riffle or mid-stream kick (if there is no riffle present) and an approximately 12-minute, 50-m riparian bank sample. The contents were elutriated six times and poured through a #30 USGS sieve. The remaining content in the sieve was then subsampled for 15 minutes. Organisms were placed in a vial with 99.5% isopropyl alcohol and returned to the lab for later identification.

Field sheets (Appendix B, Table 14) were

mIBI Sub-Metrics and Stand-Alone Indices	Response to Disturbance
Total Number of Taxa	Decrease
Total Abundance of Individuals	Decrease
Number of EPT taxa	Decrease
% Orthocladiinae & Tanytarsini	Increase
% Non-Insects (-Crayfish)	Increase
Number of Dipteran Taxa	Increase
% Intolerant Taxa (Score 0-3)	Decrease
% Tolerant Taxa	Decrease
% Predators	Decrease
% Shredders & Scrapers	Decrease
% Collectors/Filterers	Increase
% Sprawlers	Decrease
Hilsenhoff Biotic Index	Increase
Shannon-Wiener Diversity Index (H')	Decrease
Shannon Evenness Index (J')	Decrease
% Dominance of Top Three Taxa	Increase
% Chironomidae	Increase

Table 1.—mIBI submetrics and stand alone indices and their response to disturbance

completed, including the "Qualitative Habitat Evaluation Index" sheet (Appendix B, Table 18). Taxa sheets for each macroinvertebrate site can be found in Appendix B, Table 15. QHEI sheets and tabulations can be found in Appendix B, Table 18.

Macroinvertebrate Laboratory Methods.— All organisms were identified to the lowest practical level, usually genus. Non- Chironomid macroinvertebrates were identified using numerous dichotomous keys recommended in IDEM's protocol, as well as Peckarsky et al. (1990). Chironomids (with heads removed) were mounted on slides in a high viscosity mountant. Chironomids were then identified using Peckarsky et al. (1990), Mason (1998), and Epler (2001).

Macroinvertebrate Data Tabulation.— Macroinvertebrate calculations were based on IDEM's Macroinvertebrate Index of Biotic Integrity (mIBI), the Hilsenhoff Biotic Index (HBI), Shannon-Wiener Diversity Index (H'), Shannon Evenness Index (J'), Percent Dominance of Top Three Taxa, and Percent Chironomidae.

IDEM's Macroinvertebrate Index of Biotic Integrity (mIBI): The mIBI is a multimetric index (Table 1) that has been calibrated using statewide data. After calculating each metric, the resulting score is assigned a specific "rank" (1, 3, or 5) based on the drainage area of the site. The sum of all metrics is then used to determine the final score. This final score is assigned a narrative rating (Table 2). IDEM ratings also include a designation of "Fully Supporting" of aquatic life (mIBI score \leq 36), or "Not Supporting" of aquatic life (mIBI score \leq 36).

Table 2.—mIBI scores and corresponding ratings.

Narrative Rating

Excellent

Good

Fair

Poor

Very Poor

Total Score

54-60

44-53

35-43

23-34

0-22

Hilsenhoff Biotic Index (HBI): The HBI (Hilsenhoff 1987) is a biotic index that incorporates a weighted relative abundance of each taxon in order to determine a score for the community (Rosenberg & Resh 1993). Organisms are assigned a value between 0 and 10, according to their tolerance of organic and nutrient pollution (Appendix B, Table 10). The number of each organism is multiplied by the tolerance value. The sum of these results is then averaged to get the resulting HBI value for the site. Modified descriptive ratings can be found below in Table 3.

The Hilsenhoff Biotic Index is calculated as follows:

$$HBI = \sum \frac{x_i t_i}{N}$$

 X_i = number of each species

Where:

 T_i = tolerance value for each species (Appendix. B, Table 10)

N = total number of arthropods in the sample with tolerance ratings

Table 3.—HBI values and corresponding ratings.

g	HBI Score	Water Quality	Degree of Organic
) f			Pollution
l e	0.00-3.50	Excellent	No apparent organic pollu- tion.
a e c	3.51-4.50	Very Good	Possible slight organic pollution.
C	4.51-5.50	Good	Some organic pollution.
	5.51-6.50	Fair	Fairly significant organic pollution
g	6.51-7.50	Fairly Poor	Significant organic pollu- tion.
	7.51-8.50	Poor	Very significant organic pollution.
	8.51-10.00	Very Poor	Severe organic pollution.

Shannon-Wiener Diversity Index (H'): The Shannon-Wiener Diversity Index is based on the premise that species diversity decreases with decreasing water quality (Wilhm 1967; Rosenberg & Resh 1993) in an effectively infinite community (Kaesler et al. 1978). This index incorporates both species richness as well as evenness (Ludwig & Reynolds 1988). Higher H' scores indicate Tabl increased species diversity (Vandermeer 1981; ratings. Gerritsen et al. 1998). The Shannon Wiener Index is calculated as follows:

$$H' = \sum p_i \ln p_i$$

 p_i = relative abundance of each species calculated as a proportion of individuals of a given species to the total number of individuals in the community.

Shannon Evenness Index (J'): Shannon Evenness Index (Pielou 1966) is calculated from ______ the Shannon-Wiener Diversity Index and is a ratio of observed diversity to maximum diversity in order to measure evenness of the community. Higher J' scores indicate increased community evenness.

The Shannon Evenness Index is calculated as follows:

$$J' = \frac{H'}{\ln s}$$

Where:

Where:

s = number of species

Percent Dominance of Top Three Taxa: A well balanced community is indicative of a healthy community. Predominance of only a few macroinvertebrate species can be indicative of stressors in the system (Plafkin et al. 1989; Klemm et al. 1990).

Percent Chironomidae: Chironomidae are generally considered to be pollution tolerant. An overabundance of these organisms can be indicative of stressors in the system (Plafkin et al. 1989; Barbour et al. 1994).

Qualitative Habitat Evaluation Index (QHEI): The QHEI was assessed to better determine the effect of habitat quality on the resulting scores. The QHEI (Rankin 1989) is an index that evaluates macro-habitat quality that has been found to be essential for fish communities as well as other aquatic life. QHEI metrics include substrate, instream cover, channel morphology, riparian condition, pool and riffle quality, and gradient. Each metric in the habitat assessment was scored, with the final sum of these scores reflecting available habitat (higher scores reflect better habitat). Narrative ratings for QHEI scores can be found in Table 4.

Table 4.—QHEI scores and corresponding atings.

QHEI score	Narrative Rating	
90-100	Excellent	
71-89.9	Good	
52-70.9	Fair	
27-51.9	Poor	
0-26	Very Poor	
2.0		

MUSSEL RESULTS

In 2018, 120 sampler hours yielded 4,081 Unionid mussels of 17 species found within the city limits of Muncie. The most common mussels found were *Actinonaias ligamentina* (35.2%) and *Lasmigona costata* (26.2%). Most sites (30 of 37 sites) sampled in Muncie had zebra mussels present.

MACROINVERTEBRATE RESULTS

mIBI.—White River: White River mIBI scores (Graph 2 and Appendix B, Table 10) ranged from 30.0 (WHI 320.1) to 48.0 (WHI 333.4), *Poor* to *Good.* In 2018, WHI 320.1, WHI 317.6, WHI 315.0, and WHI 311.7 would be considered "Not Supporting" of aquatic life by IDEM. Mean mIBI scores (Appendix B, Table 11) upstream, within, and downstream of Muncie were all *Fair.*

Buck Creek: Buck Creek mIBI scores (Graph 3 and Appendix B, Table 10) ranged from 28.0 (BUC 13.8, BUC 5.9, and BUC 0.9) to 38.0 (BUC 9.2), *Poor* to *Fair*. The mean mIBI score for Buck Creek was 32.7, *Poor* (Appendix B, Table 11). In 2018, BUC 15.2, BUC 14.9, BUC 13.8, BUC 11.3, BUC 9.5, BUC 8.0, BUC 5.9, and BUC 0.9, and BUC 0.2 would be considered "Not Supporting" of aquatic life by IDEM. No spatial or temporal trends were detected.

In addition to the temporal trends detected from 2014-2018, a few observations should be noted. On White River, there have only been three *Poor* mIBI scores upstream of Muncie since 2009. Scores appear to fluctuate on White River from



Graph 2.—Buck Creek mIBI scores, 2018







year to year, especially dramatic in recent years. Lower mIBI scores appear to be fairly common among tributary sites.

Smaller Tributary Sites: mIBI scores at the smaller tributaries (Graph 4 and Appendix B, Table 10) ranged from 24 (MUN 2.2) to 38 (GRE 0.1) *Poor* to *Fair*. MUN 2.2, MUN 0.1, YOR 8.6, YOR 7.4, and YOR 6.3 would be considered "Not Supporting" of aquatic life by IDEM. Since 2014, mIBI scores have significantly decreased ($R^2 = 0.81$, p < 0.05) at YOR 8.6.

Stand Alone Indices.—

HBI: White River: White River HBI scores (Graph 5 and Appendix B, Table 10) ranged from 5.63 (WHI 313.4) to 3.56 (WHI 318.8), *Fair* to *Very Good*. Mean HBI scores (Appendix B, Table 11) dropped slightly from *Very Good* to *Good* within Muncie, and dropped slightly downstream of Muncie city limits. Since 2014, HBI scores have increased at WHI 317.6 ($R^2 = 0.82$, p < 0.05). No spatial or temporal trends were detected.

Buck Creek: Buck Creek HBI scores (Graph 6, Appendix B, Table 10) ranged from 6.85 (BUC 5.9) to 4.44 (BUC 9.5), *Fairly Poor* to *Very Good*. The mean HBI score (Appendix B, Table 11) was 5.6, *Fair.*). No spatial or temporal trends were detected.

Smaller Tributary Sites: HBI scores at the smaller tributaries (Graph 7 and Appendix B, Table 10) ranged from 7.65 (MUN 2.2) to 5.33 (YOR 6.3), *Poor* to *Good*.

H': White River: White River H' scores (Graph 8 and Appendix B, Table 10) ranged from 1.74 (WHI 315.0) to 3.47 (WHI 328.1 and WHI 30.7). Mean H' scores (Appendix B, Table 11) dropped as White River progressed into Muncie, but improved slightly downstream of city limits. Shannon-Wieiner scores significantly decreased since 2014 at MUN 0.1 ($R^2 = 0.91$, p < 0.05), and YOR 8.6 ($R^2 = 0.78$, p < 0.05). No spatial trends were detected.

Buck Creek: Buck Creek H' scores (Graph 9 and Appendix B, Table 10) ranged



Graph 5.—Buck Creek HBI scores, 2018.





Graph 6.—Tributary HBI scores, 2018.

from 2.39 (BUC 9.5) to 3.14 (BUC 0.0). The mean H' score at Buck Creek sites in 2018 (Appendix B, Table 11) was 2.71. No spatial or temporal trends were detected in 2018.

Smaller Tributary Sites: H' scores at the smaller tributaries ranged from (Graph 10 and Appendix B, Table 10) 1.09 (MUN 0.1) to 3.47 at GRE 0.1.

Remaining Stand Alone Indices: White **River:** White River J' scores (Appendix B, Table 10) ranged from 0.59 (WHI 315.0) to 0.91 (WHI 320.1 and WHI 318.3). Mean J' scores (Appendix B, Table 11) worsened as White River progressed downstream. White River "Percent Dominance of Top Three Taxa" (Appendix B, Table 10) ranged from 0.22 (WHI 328.1) to 0.59 (WHI 315.0). (Appendix B, Table 11) Mean scores White worsened as River progressed downstream. White River "Percent Chironomidae" (Appendix B, Table 10) ranged from 0.00 (WHI 333.4, WHI 318.8, WHI 317.2, and WHI 315.8) to 0.23 (WHI 313.4). Mean scores (Appendix B, Table 11) worsened within city limits, then improved slightly as White River progressed downstream.

Buck Creek: Buck Creek J' scores (Appendix B, Table 10) ranged from 0.73 (BUC 9.5) to 0.93 (BUC 11.3 and BUC 0.0). The mean Buck Creek J' score (Appendix B, Table 11) was 0.80. Buck Creek "Percent Dominance of Top Three Taxa" (Appendix B, Table 10) ranged from 0.57 (BUC 5.9) to 0.30 (BUC 0.0), with a mean of 0.50 (Appendix B, Table 11). Buck Creek "Percent Chironomidae" scores (Appendix B, Table 10) ranged from 0.49 (BUC 4.0) to 0.00 (BUC 15.2 and BUC 0.2), with a mean of 0.10 (Appendix B, Table 11).

Smaller Tributary Sites: J' scores at the smaller tributaries (Appendix B, Table 10) ranged from 0.51 (YOR 8.6) to 0.91 (MUN 0.1 and YOR 6.3). "Percent Dominance of Top Three Taxa" ranged from (Appendix B, Table 10) 0.58 (MUN 2.2) to 0.35 (YOR 6.3). "Percent Chironomidae" (Appendix B, Table 10) ranged from 0.06 (YOR 7.4) to 0.00 (YOR 8.6 and YOR 6.3).



Graph 8.—Buck Creek H' scores, 2018.





Graph 9.—Tributary H' scores, 2018.

OHEI: White River: White River QHEI scores ranged from 58.8 (WHI 311.7) to 82.0 (WHI 306.5), Fair to Good (Graph 11 and Appendix B, Table 10). Mean scores worsened within Muncie city limits, but recovered downstream (Appendix B, Table 11). Since 2014, OHEI scores have significantly increased at WHI 333.4 ($R^2 =$ 0.85, p < 0.05). No spatial trends were detected in 2018.

Buck Creek: Buck Creek OHEI scores (Graph 12 and Appendix B, Table 10) ranged from 39.5 (BUC 11.3) to 73.0 (BUC 0.0), Poor to Good, with a mean score of 58.1, Fair (Appendix B, Table 11). Since 2014, QHEI scores significantly increased at BUC 0.0 ($R^2 = 0.82$, p < 0.05). No spatial trends were detected in 2018.

Smaller Tributary Sites: QHEI scores at the smaller tributaries ranged from (Graph 13 and Appendix B, Table 10) 43.8 (YOR 8.6) to 66.5 (YOR 6.3), Poor to Fair. Since 2014, QHEI scores have significantly decreased at YOR 8.6 ($R^2 = 0.92$, p < 0.05), and have significantly increased at YOR 6.3 ($R^2 =$ 0.98, p < 0.01). No spatial trends were detected in 2018.

DISCUSSION

Mussels.—Mussel sampling results continue to indicate good water quality within Muncie city limits, considering the urban impacts on this stretch of White River.

It has been noted that one mussel species, the white heelsplitter Lasmigona complanata, has not been found in White River upstream of Muncie. This species' opportunistic nature, and its ability to tolerate silt, habitat disturbance, and impoundments (Grabarkiewicz & Davis 2008), appear to make it an ideal species to inhabit White River within city limits. However, it is possible that this species is unable to expand its range upstream due to the inability of its species to navigate the five host impoundments within Muncie city limits.

Dams are well documented as obstacles to mussel population abundance and expansion (Vaughn & Taylor 1999; Watters 2000; Dean



Graph 11.—Buck Creek QHEI scores, 2017.





Graph 132—Tributary QHEI scores, 2017.

et al. 2002). Habitats are altered upstream and downstream of the impoundment, resulting in an increase of pollutants, siltation, stagnation, thermal changes, and anoxic conditions (Watters 1999), causing additional complications for mussel populations (Watters 1996; Dean et al. 2002; Lessard & Haves 2003; Tienmann et al. 2004; Poff et al. 2007; Maloney et al. 2008). Dams have been implicated as one of the leading causes of current-day decline in freshwater mussel populations in North America (Parmalee & Bogan 1998; Haag 2009). They have been cited as being responsible for the "local extirpation of 30-60% of the native freshwater mussel species in many United States rivers" (NRCS 2009). Studies have shown that the impacts of impoundments have resulted in reduced abundance, diversity, and species richness of mussel fauna (Dean et al. 2002; Baldigo et al. 2004; Tiemann et al. 2004; Santucci et al. 2005; Galbraith & Vaughn 2011: Tiemann et al. 2016).

In late summer 2017, zebra mussels were found in White River downstream of Prairie Creek Reservoir (where they were first observed in 2015). Within weeks, zebra mussels were identified on dead mussel shell in the WR 313.4 site. In 2018, random quadrat sampling at each macroinvertebrate site yielded densities of 0-36/m² at sites between WHI 322.2-WHI 308.7.

Timed search surveys will continue in 2019, likely upstream of Muncie, and will continue until all of the West Fork of White River in Delaware County has been assessed.

Macroinvertebrates—Many sites had lower mIBI scores in 2018. Most of these sites also had unusually low abundance and/or diversity.

Poor mIBI scores at some sites may be attributed to a lack of suitable habitat for macroinvertebrates, quantified by *Poor* QHEI scores. Sites at BUC 11.3, BUC 9.5, BUC 8.0, GRE 0.1, MUN 2.2, YOR 8.6, and YOR 7.4 all had *Poor* QHEI scores, indicating that a lack of habitat may limit the macroinvertebrates that can inhabit these sites.

Organic impairment appears to be a likely stressor at one site. BUC 5.9 is the only site in 2018 to have a *Fairly Poor* HBI score., coinciding with a *Poor* mIBI as well.

Many remaining sites with *Poor* mIBI scores do not suggest organic impairment or habitat limitations. Most of these sites have very low abundance and/or diversity, exaggerating any effects on this sample and carrying over into multiple metrics. These include BUC 15.2, BUC 14.9, BUC 13.8, BUC 0.9, BUC 0.2, BUC 0.0, MUN 0.1, WHI 320.1, WHI 315.0, WHI 311.7, and YOR 6.3..

Only WHI 317.6 had a *Poor* mIBI, but did not have low abundance. This site was dominated (50%) by *Hyallela azteca*, *Goniobasis livascens*, and *Caenis* spp. *H. azteca* and *G. livascens* are moderately tolerant to tolerant non-insects, and their dominance affects multiple indices. This site was also dominated (49.8%) by non-insects, and had 37.0% tolerant organisms.

Significant decreases in mIBI scores from 2014-2018 indicate potential water quality issues at YOR 8.6 (*Fair* to *Poor*). Conversely, a significant increase in mIBI scores from 2014-2018 was seen at WHI 333.4 (*Fair* to *Good*), indicating potentially improved water quality.

A significant increase in HBI scores from 2014-2018 at WHI 317.6 suggests decreased water quality, specifically increased organic enrichment, at this site.

Significant decreases in H' scores from 2014-2018 show decreased diversity in macroinvertebrate populations at some sites, potentially indicating stressors at these sites. These sites include MUN 0.1 (3.35-1.09) and YOR 8.6 (2.31-1.35).

Significant increases in QHEI scores from 2014-2018 indicate increased habitat availability at some sites. These sites include WHI 333.4 (*Fair* to *Good*), BUC 0.0 (*Fair* to *Good*), and YOR 6.3 (*Poor* to *Fair*). Significant decreases in QHEI scores from 2014-2018 indicate decreased habitat availability at YOR 8.6 (*Fair* to *Poor*).

Observed trends give us some indication of negative impacts on sample sites. *Poor* mIBI scores (as seen in 2018 at WHI 320.1) generally are not seen on White River upstream of Muncie city limits, and is unusual for this site. Scores from

this site will be watched closely in future years to determine if this is indicative of a trend.

Multiple negative mIBI scores at tributary sites likely reflect impacts that are more apparent due to their smaller size. Additionally, diversity and/or abundance may be limited by the colder temperatures found in spring-fed Buck Creek (Vannote & Sweeney 1980; Ward 1976).

Climatological fluctuations and extremes have been considered as factors in years with unusually low mIBI scores (Bowley 2012; Bowley 2015; Bowley 2016). Other stressors may need to be considered including the effects of multiple stressors. These may include ecological. morphological, hydrological, biological, chemical or climatological effects. To complicate an already challenging situation. most aquatic macroinvertebrates have complex life cycles that include multiple stages, some being terrestrial.

An emerging global concern has also been considered for the recent drop in scores, particularly in abundance and diversity. А growing body of evidence has supported what is being called an "Insect Apocalypse", indicating an alarming drop in insect abundance and diversity worldwide. A study in Germany's protected areas found a 76% seasonal decline in insect biomass of over 27 years (Hallmann et al. 2017), finding no significant correlation with landuse, habitat or climate change. A study in Puerto Rico showed a 2.2-2.7% annual loss in ground-dwelling and canopy-dwelling arthropods (Lister & Garcia 2018), indicating "climate warming" as the likely cause. Similar declines in flying insects have been seen in areas all around the globe (Thomas et al. 2004; Shortall et al. 2009; Sanchez-Bayo & Wyckhuys 2019).

Since most flying insects spend part of their life cycle as aquatic insects, it stands to reason that a similar trend would be seen at an aquatic level. Declines in abundance and diversity, and increases in homogeneity and/or replacement from tolerant and generalist species has been seen in the Odonata (Hickling et al. 2005; McKinney 2006; Kadoya et al. 2009; Kalkman et al. 2010), Ephemeroptera (Zahradkova et al. 2009; Zedkova et al. 2015), and Trichoptera orders (Karatayev et al. 2009; Houghton & Holzenthal 2010; Jenderedjian et al. 2012). Future work at the BWQ will be looking at long-term trends in our macroinvertebrate data to determine if sites in this area are experiencing similar trends. Research and analysis, as well as continued monitoring, will be conducted in an attempt to determine all stressors affecting macroinvertebrate communities.

Dramatic improvements have been seen since the inception of our macroinvertebrate and mussel sampling programs. Point source pollutants have been controlled through the utilization of local permits regulated by the Bureau of Water Quality. Improvements have been and continue to be made to our Water Pollution Control Facility. Whereas most analyses historically have focused on White River, studying the tributaries and nonpoint source pollution impacting them has become critical. These impacts on water quality include hydromodifications (channelization, impoundments, dredging, and removal of riparian zones), urban storm water (sources include CSOs, SSOs. and impervious surfaces). and sedimentation. In 1990, the US EPA listed sedimentation as the top pollutant of rivers in the United States (Box & Mossa 1999), and it has been determined that reductions in water quality are detectable at > 15% impervious surface (Roy et al. 2003).

This shift in focus would benefit from public outreach, education, and cooperation to instill better management practices throughout Delaware County. These include buffer strips, rain barrels, rain gardens, better construction site practices, and the further separation of CSOs. As improved management practices are implemented, it is expected that water quality will continue to improve.

Overall, the water systems in this area appear to be in good condition, especially considering the industrial, urban, and agricultural areas through which they flow. Efforts by the citizens of Delaware County, the City of Muncie, the Muncie Sanitary District, the Bureau of Water Quality, and the industrial community are responsible for the improvements in water quality since the BWQ was established in 1972. Appendix A.—Mussel assemblages and relative abundance found within city limits, 2018.

Table 4.—Mussel assemblage within Muncie city limits, 2018.

Scientific Name	Common Name	# Found
Lasmigona costata	flutedshell	1068
Actinonaias ligamentina	mucket	1437
Lampsilis siliquoidea	fatmucket	330
Lasmigona complanata	white heelsplitter	271
Alasmidonta marginata	elktoe	217
Amblema plicata	threeridge	211
Lampsilis cardium	plain pocketbook	178
Fusconcaia flava	Wabash pigtoe	133
Pleurobema sintoxia	round pigtoe	54
Eurynia dilatata	spike	50
Strophitus undulatus	creeper	45
Lampsilis fasciola	wavy-rayed lampmussel	25
Villosa iris	rainbow	24
Pyganodon grandis	giant floater	20
Anodontoides ferussacianus	cylindrical papershell	7
Lasmigona compressa	creek heelsplitter	6
Utterbackia imbecillis	paper pondshell	5
	TOTAL	4081

Graph 13.—Relative abundance for Unionid mussels sampled within Muncie city limits,, 2018.



Appendix B.—Macroinvertebrate sites, field sheets, tolerance and attributes used for calculations, taxa identified, taxa sheets, QHEI sheets, and resulting scores.

Table 5.—Macroinvertebrate site field sheets, 2018.

BUREAU OF WATER QUALITY MUSSEL BED SURVEY

Stream	Station	County	Date	
Collected by:				
Collection Notes:				
Width:				
1	26	51	76	
2	20	52	77	
3	28	53	78	
4	29	53	79	
5	30	55	80	
6	31	56	81	
7	32	57	82	
8	33	58	83	
9	34	59	84	
10	35	60	85	
11	36	61		

8	33	58	83	
9	34	59	84	_
10	35	60	85	_
11	36	61	86	_
12	37	62	87	
13	38	63	88	_
14	39	64	89	
15	40	65	90	_
16	41	66	91	
17	42	67	92	_
18	43	68	93	_
19	44	69	94	_
20	45	70	95	_
21	46	71	96	
22	47	72	97	_
23	48	73	98	_
24	49	74	99	
25	50	75	100	

Table 5.—Macroinvertebrate site field sheets, 2018 (con't).

Bureau of Water Quality Mussel Data

Stream		Station	Date			
Transect	Collector	Species	Width	Height	Age	Count
					- 3-	
					_	
					_	
	-					
	-					
					_	
					_	
			I			
						_
					_	-
			<u> </u>		_	-
						_

Buck Creek	CR 950N (BUC 15.2)	Lat./Long.	40.070817	-85.363497
Drainage=13 sq.miles	HUC14: 05120201020020			
Water is much colder (4.2°C to	o 6.5°C low er than White River) due to the sy	stem being spring fed	(Conrad and War	rner 2005).
Buck Creek	CR 800S (BUC 14.9)	Lat./Long.	40.076306	-85.362624
Drainage= 27 sq. miles	HUC14: 05120201020020			
Water is much colder (4.2°C to	o 6.5°C low er than White River) due to the sy	stem being spring fed	(Conrad and War	rner 2005).
Buck Creek	CR 700S (BUC 13.8)	Lat./Long.	40.090910	-85.361338
Drainage= 27 sq. miles	HUC14: 05120201020020			
Water is much colder (4.2°C to	o 6.5°C low er than White River) due to the sy	stem being spring fed	(Conrad and War	rner 2005).
Buck Creek	SR 3 (BUC 11.3)	Lat./Long.	40.123676	-85.370897
Drainage= 36 sq. miles	HUC14: 05120201020020			
Water is much colder (4.2°C to	o 6.5°C low er than White River) due to the sy	stem being spring fed	(Conrad and War	rner 2005).
Buck Creek	CR 300S/Fuson Rd. (BUC 9.5)	Lat./Long.	40.149185	-85.378202
Drainage= 49 sq. miles	HUC14: 05120201020020			
Water is much colder (4.2°C to	o 6.5°C low er than White River) due to the sy	stem being spring fed	(Conrad and War	rner 2005).
Buck Creek	Madison St. (BUC 9.2)	Lat./Long.	40.155806,	-85.382286
Drainage= 49 sq. miles	HUC 14: 05120201020020	-		
Water is much colder (4.2°C to	o 6.5°C low er than White River) due to the sy	stem being spring fed	(Conrad and War	rner 2005).
Buck Creek	23rd St. (BUC 8.0)	Lat./Long.	40.16756,	-85.391803
Drainage= 49 sq. miles	HUC 14: 05120201020020	-		
Water is much colder (4.2°C to	o 6.5°C low er than White River) due to the sy	stem being spring fed	(Conrad and War	rner 2005).
Buck Creek	Tillotson Ave. (BUC 5.9)	Lat./Long.	40.174127	-85.423697
Drainage= 49 sq. miles	HUC14: 05120201020020	-		
•	o 6.5°C low er than White River) due to the sy	stem being spring fed	(Conrad and War	rner 2005).
Buck Creek	CR 325W (BUC 4.0)	Lat./Long.	40.15686,	-85.446570
Drainage= 49 sq. miles	HUC 14: 05120201020060			
•	o 6.5°C low er than White River) due to the sy	stem being spring fed	(Conrad and War	rner 2005).
Buck Creek	Cornbread Rd. W. Crossing (BUC 0.9)	Lat./Long.	40.170817	-85.487403
Drainage= 100 sq. miles	HUC 14: 05120201020060			
•	0 6.5°C low er than White River) due to the sy	stem being spring fed	(Conrad and War	rner 2005).
Buck Creek	SR 32 (BUC 0.2)	Lat./Long.	40.174756,	-85.493202
Drainage= 100 sg. miles	HUC 14: 05120201020060	Ū		
0 1	o 6.5°C low er than White River) due to the sy	stem beina sprina fed	(Conrad and War	rner 2005).
Buck Creek	Confluence (BUC 0.0)	Lat./Long.	40.174082,	-85.500697
Drainage= 100 sq. miles	HUC 14: 05120201020060		,	
•	merous band stabilization efforts, this site uno	derwent reconstructio	n in the fall of 201	3. This site was
	013, and will be sampled annually hereafter to			
• •	ed, and large boulders and j-hooks were insta			•
•	0.5° C low er than White River) due to the sy	-		-
20. Greenfarm Ditch	Moore Rd. (GRE 0.1)	Lat./Long.	40.236342,	-85.414939
Drainage=3 sq. miles		Eut./Long.		
•	is site is primarily residential and commercial.	Both banks are mowe	ed to the edge	
This site is just within the influ			sa to the edge.	
Muncie Creek	Indiana Ave. (MUN 2.2)	l at // ong	40.226458,	-85.361522
Drainage= 10.0 sq. miles	HUC 14: 05120201010130	Lat./Long.	,	
Muncie Creek		l at // ong	40.201933,	-85.379461
	McCulloch Park (MUN 0.1)	Lat./Long.		00.010101
Drainage= 10.0 sq. miles	HUC 14: 05120201010130	Lat // one	40 165022	05 100040
West Fork White River	CR 1100W (WHI 333.4)	Lat./Long.	40.165932,	-85.182243
Drainage= 120 sq. miles	HUC 14: 05120201010090	1 -4 "	40.405050	05 050040
West Fork White River	CR 700E (WHI 328.1)	Lat./Long.	40.165859,	-85.253616
Drainage= 184 sq. miles	HUC 14: 05120201010100			

Table 6.—Macroinvertebrate site descriptions and locations, 2018.

West Fork White River	Smithfield (WHI 326.9)	Lat./Long.	40.168793,	-85.271332
Drainage= 184 sq. miles	HUC 14: 05120201010100			
West Fork White River	Camp Red Wing (CRW) (WHI 322.2)	Lat./Long.	40.145227,	-85.322876
Drainage= 213 sq. miles	HUC 14: 05120201010120			
West Fork White River	Burlington (WHI 320.1)	Lat./Long.	40.169697,	-85.341393
Drainage= 220 sq. miles	HUC 14: 05120201010120			
Large man-made boulder and	cobble riffle stretches the width of the stream.			
West Fork White River	Water Company (WHI 318.8)	Lat./Long.	40.183727,	-85.349831
Drainage= 220 sq. miles	HUC 14: 05120201010120			
Site dow nstream of Water Co	mpany low head dam. Riffle sampled in riffle and	dam for consisten	cy to past efforts.	
West Fork White River	River Rd. (WHI 318.3)	Lat./Long.	40.184911,	-85.429108
Drainage= 220 sq. miles	HUC 14: 05120201010120			
West Fork White River	E Jackson (WHI 317.6)	Lat./Long.	40.194584,	-85.364861
Drainage= 231 sq. miles	HUC 14: 05120201010130			
Site substrate almost exclusiv	ely bedrock.			
West Fork White River	Bunch Blvd. (WHI 317.2)	Lat./Long.	40.198117,	-85.367828
Drainage= 231 sq. miles	HUC 14: 05120201010130			
West Fork White River	Em St. (WHI 315.8)	Lat./Long.	40.204031,	-85.386483
Drainage= 241 sq. miles	HUC 14: 05120201020060			
Substrate is dominated by bec	Irock.			
West Fork White River	High St. (WHI 315.0)	Lat./Long.	40.195446,	-85.390610
Drainage= 241 sq. miles	HUC 14: 05120201020060			
Site is dow nstream of large lo	w head dam in dow ntow n Muncie.			
West Fork White River	Tillotson Ave. (WHI 313.4)	Lat./Long.	40.184975,	-85.421722
Drainage= 245 sq. miles	HUC 14: 05120201020060			
West Fork White River	Above MWWPCF (WHI 311.7)	Lat./Long.	40.185396,	-85.439118
Drainage= 245 sq. miles	HUC 14: 05120201020060			
West Fork White River	CR 400W/Nebo Rd. (WHI 310.7)	Lat./Long.	40.186045,	-85.462912
Drainage= 246 sq. miles	HUC 14: 05120201020060			
This is the first annual baseline	e site dow nstream of the MWPCF.			
West Fork White River	CR 575W (WHI 308.7)	Lat./Long.	40.177713,	-85.497803
Drainage= 248 sq. miles	HUC 14: 05120201020060			
West Fork White River	CR 750W (WHI 306.5)	Lat./Long.	40.165253,	-85.530273
Drainage= 367 sq. miles	HUC 14: 05120201030010			
Flow is extremely fast at this	site.			
West Fork White River	CR 300S (WHI 304.4)	Lat./Long.	40.148876,	-85.552838
Drainage= 370 sq. miles	HUC 14: 05120201030020			
Flow is very fast at this site.				
York Prairie Creek	Brook Rd./Storer Elem. (YOR 8.6)	Lat./Long.	40.206286,	-85.423686
Drainage= 4.00 sq. miles	HUC 14: 05120201030010			
York Prairie Creek	CR 300W (YOR 7.4)	Lat./Long.	40.199781,	-85.443308
Drainage= 4.00 sq. miles	HUC 14: 05120201030010			
York Prairie Creek	CR 400W (YOR 6.3)	Lat./Long.	40.193758,	-85.460747
Drainage= 4.00 sg. miles	HUC 14: 05120201030010			

Table 6.—Macroinvertebrate site descriptions and locations, 2018 (con't).

Species	Tolerance Value
Ablabesmyia	5
Ablabesmyia annulata	4
Ablabesmyia janta	5
Ablabesmyia mallochi	5
Acariformes	4
Acentrella	4
Acentrella ampla	6
Acentria	5
Acerpenna	4
Acerpenna macdunnoughi	1
Acerpenna pygmaea	2
Acroneuria	1
Acroneuria abnormis	0
Acroneuria evoluta	3
Acroneuria internata	2
Acroneuria lycorias	2
AESHNIDAE	3
Agabetes	5
Agabus	5
Agapetus	0
Agnetina	2
Agnetina annulipes	2
Agnetina capitata	2
Agnetina flavescens	2
Agraylea	6
Allocapnia	3
Allocapnia vivipara	3
Alloperla	0
Ameletus	0
Ameletus lineatus	0
Ameletus ludens	0
AMNICOLA	5
Amnicola limosus	5
Amphinemura	3
Amphinemura delosa	3
Amphinemura nigritta	3
AMPHIPODA	4
ANCYLIDAE	6
Ancyronyx variegatus	4
Anthopotamus	4
Anthopotamus verticis	4
Antocha	2
Arcteonais lomondi	6
Argia	5
ASELLIDAE	8
ASTACIDAE	6
ATHERICIDAE	2
Atractides	6
Atrichopogon	5
Atrichopogon websteri	4

Species	Tolerance Value
Attenella attenuata	3
Aulodrilus	7
Aulodrilus americanus	7
Aulodrilus limnobius	7
Aulodrilus pigueti	7
Aulodrilus pluriseta	7
BAETIDAE	4
Baetis	3
Baetis brunneicolor	4
Baetis flavistriga	3
Baetis intercalaris	3
Baetis tricaudatus	4
Baetisca	4
BAETISCIDAE	3
Basiaeschna	6
Basiaeschna janata	6
Belostoma flumineum	4
Berosus	7
Berosus peregrinus	6
Berosus striatus	5
BITHYNIA	8
Bithynia tentaculata	8
BLEPHARICERIDAE	0
vejdovskyanum	° 7
Boyeria	2
Boyeria vinosa	4
BRACHYCENTRIDAE	1
Brachycentrus lateralis	1
Brachycentrus numerosus	1
Brachycercus	3
BRANCHIOBDELLIDAE	6
Branchiura	6
Branchiura sowerbyi	6
Brillia	5
Caecidotea	8
Caecidotea communis	8
CAENIDAE	7
Caenis	3
Callibaetis	6
Calopteryx	4
Cambarus	2
Cambarus diogenes	6
CAPNIIDAE	1
Cardiocladius	5
Cardiocladius obscurus	2
Centroptilum	3
Ceraclea	3
Ceraclea ancylus	3
Ceraclea maculata	4
CERATOPOGONIDAE	6
	v

Table 7.—Tolerance values used in mIBI/HBI calculations.

Species	Tolerance Value
Ceratopsyche alhedra	3
Ceratopsyche bronta	5
Ceratopsyche morosa	2
Ceratopsyche slossonae	2
Ceratopsyche sparna	3
Chaetogaster	7
Chaetogaster diaphanus	6
Chaetogaster diastrophus	6
Chaetogaster limnaei	6
Chaoborus	8
Chauliodes	4
Cheumatopsyche	3
Chimarra	4
Chimarra aterrima	2
Chimarra obscura	4
Chimarra socia	2
CHIRONOMIDAE(all other)	6
CHIRONOMIDAE(blood red)	8
Chironomus	8
CHLOROPERLIDAE	1
Choroterpes	4
Chrysops	5
Cincinnatia cincinnatiensis	5
Cladopelma	9
Cladotanytarsus	4
Climacia	5
Clinotanypus pinguis	8
Clioperla clio	1
Cloeon	4
Cnephia mutata	5
COENAGRIONIDAE	9
Conchapelopia	4
Corbicula fluminea	6
Cordulegaster	3
CORDULEGASTRIDAE	3
CORDULIIDAE	3
CORIXIDAE	5
CORYDALIDAE	1
Corydalus cornutus	2
Corynoneura	4
Corynoneura celeripes	2
Crangonyx	6
Crenitis	5
Cricotopus	5 4
Cricotopus bicinctus	4 7
Cryptochironomus	7 5
Cryptochironomus blarina	5 8
Cryptochironomus fulvus	8
	8 4
Cryptotendipes CULICIDAE	4 8
	U

Species	Tolerance Value
Culicoides	10
CURCULIONIDAE	5
Cyrnellus fraternus	4
Dannella	2
Dannella lita	4
Dero	10
Dero digitata	10
Dero furcata	10
Dero nivea	10
Dero obtusa	10
Dero vaga	10
Diamesa	8
Dibusa angata	3
Dicranota	3
Dicrotendipes	6
Dicrotendipes fumidus	6
Dicrotendipes modestus	6
Dicrotendipes neomodestus	5
Dineutus	4
Dineutus assimilis	4
Dineutus horni	4
Dineutus nigrior	4
Diplocladius cultriger	8
Dixa	1
DOLICHOPODIDAE	4
Dolophilodes	0
Doncricotopus bicaudatus	5
Dreissena polymorpha	8
Dromogomphus	6
Drunella walkeri	0
DRYOPIDAE	5
Dubiraphia	5
Dubiraphia bivittata	3
Dubiraphia guadrinotata	3
Eccoptura	3
Eclipidrilus	5
Ectopria	5
Ectopria nervosa	4
Elliptio complanata	8
ELMIDAE	4
EMPIDIDAE	6
Enallagma	9
ENCHYTRAEIDAE	10
Endochironomus	6
Endochironomus nigricans	5
Epeorus	0
Ephemera	3
Ephemerella	3
Ephemerella dorothea	1
Ephemerella excrucians	1

Table 7.—Tolerance values used in mIBI/HBI calculations (con't).

Species	Tolerance Value
Ephemerella invaria	1
Ephemerella needhami	2
Ephemerella subvaria	1
EPHEMERELLIDAE	1
EPHEMERIDAE	4
Ephoron	2
Ephoron leukon	2
EPHYDRIDAE	6
Erythemis	2
Eukiefferiella claripennis	8
Eurylophella	2
Eurylophella bicolor	1
Eurylophella funeralis	2
Eurylophella temporalis	5
Ferrissia	6
Ferrissia parallelus	6
Ferrissia rivularis	6
Ferrissia walkeri	6
Fossaria	6
GAMMARIDAE	4
Gammarus	6
Gammarus fasciatus	6
Gammarus pseudolimnaeus	4
GASTROPODA	7
Glossosoma	0
GLOSSOSOMATIDAE	0
Glyptotendipes	6
Goera	3
GOMPHIDAE	1
Gomphus	5
Goniobasis	6
Goniobasis livescens	6
Gyraulus	8
Gyraulus circumstriatus	8
Gyraulus deflectus	8
Gyraulus parvus	8
Gyrinus	4
Haemonais waldvogeli	8
Hagenius brevistylus	1
Haliplus	6
Haliplus borealis	5
Haliplus connexus	6
Haliplus cribrarius	6
Haliplus immaculicollis	6
Haliplus longulus	6
Haliplus pantherinus	6 1
Haploperla brevis	-
HAPLOTAXIDAE	5
Harnischia Harnischia curtilamellata	8 4
	7

Species	Tolerance Value
Helichus	5
Helichus striatus	2
Helicopsyche borealis	3
HELICOPSYCHIDAE	3
Helisoma	6
Helisoma anceps	6
Helius	4
Helobdella	10
Helobdella stagnalis	8
Helobdella triserialis	8
Helochares	5
Helophorus	5
Heptagenia	3
Heptagenia diabasia	2
Heptagenia flavescens	4
Heptagenia pulla	4
HEPTAGENIIDAE	4
Hesperocorixa	5
Hesperocorixa interrupta	5
Hesperocorixa lucida	5
Hesperocorixa vulgaris	5
Hetaerina	3
Heterocloeon	3
	2
Heterocloeon curiosum	2 0
Heterotrissocladius	
Hexagenia	4
Hexagenia limbata	3
Hexatoma	2
HIRUDINEA	8
Hyalella azteca	8
Hydatophylax	2
Hydrobaenus	8
HYDROBIIDAE	7
Hydrobius	5
Hydrobius fuscipes	4
Hydrochara	5
Hydrochus	5
Hydroporus	4
Hydropsyche	4
Hydropsyche betteni	6
Hydropsyche bidens	3
Hydropsyche depravata	6
Hydropsyche dicantha	4
Hydropsyche frisoni	2
Hydropsyche orris	3
Hydropsyche phalerata	1
Hydropsyche scalaris	2
Hydropsyche simulans	2
Hydropsyche valanis	3 3
Hydropsyche venularis	3

Table 7.—Tolerance values used in mIBI/HBI calculations (con't).

Table 7.—Tolerance values used in mIBI/HBI calculations (con't).

Species	Tolerance Valu
HYDROPSYCHIDAE	4
Hydroptila	3
Hydroptila albicornis	6
Hydroptila armata	6
Hydroptila consimilis	6
Hydroptila hamata	6
Hydroptila spatulata	6
Hydroptila waubesiana	6
HYDROPTILIDAE	4
llybius biguttulus	8
Ilyodrilus templetoni	10
Ischnura	9
Isochaetides freyi	8
Isonychia	2
Isonychia bicolor	2
ISONYCHIIDAE	2
Isoperla	2
Isoperla dicala	2
Isoperla frisoni	2
Isoperla namata	2
ISOPODA	8
Isotomurus	5
Labrundinia	4
Labrundinia pilosella	3
Laccobius	2
Laccobius spangleri	4
Laccophilus	8
Laccophilus maculosus	8
maculosus	8
Lampsilis radiata radiata	6
Larsia	4
Lebertia	4
Lepidostoma	1
LEPIDOSTOMATIDAE	1
LEPTOCERIDAE	4
Leptocerus americanus	4
Leptophlebia	4
LEPTOPHLEBIIDAE	2
Leucrocuta	2
Leucrocuta aphrodite	1
Leucrocuta hebe	3
Leucrocuta maculipennis	2
Leuctra	0
Leuctra ferruginea	0
Leuctra tenuis	0
LEUCTRIDAE	0
LEUCIRIDAE Libellula	
	9
	9
	4 3
Limnephilus	3

Species	Tolerance Value
Limnodrilus cervix	10
Limnodrilus claparedianus	10
Limnodrilus hoffmeisteri	10
Limnodrilus profundicola	10
Limnodrilus udekemianus	10
Limnophila	3
Limonia	6
Liodessus affinis	6
Liodessus flavicollis	6
Lirceus	8
LUMBRICULIDAE	5
Lutrochus laticeps	3
Lymnaea	6
adpressa	6
LYMNAEIDAE	6
Lype diversa	3
Maccaffertium exiguum	2
Maccaffertium luteum	4
mediopunctatum	2
integrum	3
Maccaffertium modestum	1
Maccaffertium pudicum	2
Maccaffertium pulchellum	2
Maccaffertium terminatum	2
Maccaffertium vicarium	2
Macromia	2
MACROMIIDAE	3
Macronychus glabratus	3
Macrostemum	3
Macrostemum carolina	3
Macrostemum zebratum	2
METRETOPODIDAE	2
Micrasema rusticum	2
Microcylloepus pusillus	3
Micropsectra	4
Microtendipes	7
Microtendipes caelum	3
Molanna	6
Molanna blenda	4
MOLANNIDAE	6
MUSCIDAE	6
Musculium	6
Musculium partumeium	6
Musculium transversum	6
Mystacides	4
Mystacides sepulchralis	4
NAIDIDAE	8
Nais	8
Nais barbata	8
Nais behningi	6

Species	Tolerance Value
Nais bretscheri	6
Nais communis	8
Nais elinguis	10
Nais pardalis	8
Nais simplex	6
Nais variabilis	10
Nanocladius	5
Nanocladius distinctus	6
Nanocladius spiniplenus	4
Natarsia	6
Natarsia baltimoreus	6
Nectopsyche	2
Nectopsyche diarina	3
	3
Nectopsyche exquisita	2
Nectopsyche pavida	
NEMATODA	6
Nemoura	1
NEMOURIDAE	2
Neoperla	3
Neophylax	3
Neophylax concinnus	3
Neophylax fuscus	3
Neotrichia	4
Neureclipsis	3
Neurocordulia obsoleta	0
Nigronia fasciatus	2
Nigronia serricornis	4
Nilotanypus	6
Nilotanypus fimbriatus	3
Nilothauma	3
Nixe	3
Nixe perfida	5
Nyctiophylax	3
Nyctiophylax moestus	5
Nymphula	7
Ochrotrichia	2
ODONTOCERIDAE	0
Oecetis	3
OLIGOCHAETA	8
OLIGONEURIIDAE	2
Oligostomis	2
Ophidonais serpentina	6
Ophiogomphus	1
Optioservus	4
Optioservus fastiditus	2
Optioservus trivittatus	4
Orconectes	4
Orconectes propinquus	4
Orconectes rusticus	6
Orconectes virilis	6
-	-

Species	Tolerance Value
Orthocladius	4
Orthocladius carlatus	2
Orthotrichia	6
Oulimnius	4
Oulimnius latiusculus	4
Oxyethira	5
Pagastia	1
Palmacorixa	5
Palmacorixa buenoi	4
Palmacorixa gillettei	4
Palmacorixa nana	4
Paracapnia	1
Paracapnia angulata	1
Parachironomus	4
Parachironomus carinatus	5
Parachironomus frequens	4
Paracladopelma	7
Paragnetina	2
0	2
Paragnetina media Parakiefferiella	2 5
	3
Paraleptophlebia	3 1
Paraleptophlebia guttata	1
Paraleptophlebia moerens	•
Paraleptophlebia mollis	1
Paraleuctra	0
Parametriocnemus	3
lundbeckii	5
Paranais frici	10
Paraponyx	5
Paratanytarsus	4
Paratendipes	6
Paratendipes albimanus	4
Pedicia	4
Pelocoris femoratus	4
Peltodytes	7
Peltodytes edentulus	6
Peltodytes tortulosus	6
Pentaneura	6
Pentaneura inconspicua	5
Pericoma	6
Perlesta	4
Perlesta placida	5
PERLIDAE	1
Perlinella drymo	1
PERLODIDAE	2
Petrophila	5
Phaenopsectra	7
Phaenopsectra flavipes	6
Phaenopsectra punctipes	4
PHILOPOTAMIDAE	3

Table 7.—Tolerance values used in mIBI/HBI calculations (con't).

Species	Tolerance Value
PHRYGANEIDAE	4
Phylocentropus	4
Physa	8
Physella	8
Physella gyrina	8
Physella heterostropha	8
Physella integra	8
PHYSIDAE	8
Pilaria	7
PISIDIIDAE	8
Pisidium	6
Pisidium casertanum	6
Pisidium compressum	6
Pisidium variabile	6
Placobdella montifera	8
PLANORBIDAE	6
Plathemis lydia	8
Platycentropus	4
Plauditus	4
Plauditus punctiventris	2
Pleurocera acuta	6
PLEUROCERIDAE	6
POLYCENTROPODIDAE	6
Polycentropus	3
POLYMITARCYIDAE	2
Polypedilum aviceps	2
Polypedilum convictum	4
Polypedilum illinoense	7
Polypedilum ontario	3
POTAMANTHIDAE	4
Potamothrix moldaviensis	8
Potamothrix vejdovskyi	8
Potamyia	5
Potamyia flava	3
Pristina	8
Pristina aequiseta	8
Pristina breviseta	8
Pristina leidyi	8
Pristina synclites	8
Pristinella	8
Pristinella jenkinae	8
Pristinella osborni	8
Probythinella lacustris	8
Procladius	7
Prodiamesa olivacea	3
Prostoia	2
Protoplasa	3
Protoptila	1
Protopilia Psectrocladius	6
Psectrotanypus	8
	0

Table 7.—Tolerance values used in mIBI/HBI calculations (con	't)
Tuble 7. Tolefallee values used in hilds/Tibl calculations (con	U.

Species	Tolerance Value
Psectrotanypus dyari	9
PSEPHENIDAE	4
Psephenus	4
Psephenus herricki	4
Pseudochironomus	5
Pseudocloeon	2
Pseudocloeon dardanus	2
Pseudocloeon propinquus	1
Pseudolimnophila	2
Pseudostenophylax	0
Pseudosuccinea columella	6
Psychoda	4
PSYCHODIDAE	10
Psychomyia flavida	2
PSYCHOMYIIDAE	2
PTERONARCYIDAE	0
Pteronarcys	0
Pteronarcys dorsata	0
Ptilostomis	5
Pycnopsyche	3
Pyganodon cataracta	6
PYRALIDAE	5
Quistradrilus multisetosus	10
Radix auricularia	6
Ranatra fusca	4
Ranatra nigra	4
Rheocricotopus	5
Rheocricotopus robacki	4
Rheotanytarsus	3
Rhithrogena	0
Rhyacodrilus	10
Rhyacophila	1
Rhyacophila glaberrima	1
RHYACOPHILIDAE	0
Ripistes parasita	8
Saetheria tylus	4
SCIRTIDAE	5
SERICOSTOMATIDAE	3
Serratella	1
Serratella deficiens	2
Setodes	2
Shipsa rotunda	2
SIALIDAE	4
Sialis	5
Sigara alternata	4
Sigara grossolineata	4
Sigara mathesoni	4
Sigara modesta	4
Sigara signata	4
Sigara variabilis	4
~	

SIMULIIDAE6Simulium5Simulium venustum5Simulium vittatum7SIPHLONURIDAE7Siphlonurus4Siphloplecton2Slavina appendiculata6Somatochlora1Sperchon4Sphaerium6Sphaerium striatinum6Spirosperma ferox6catascopium6Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenalerinis bicarinata5Stenelmis bicarinata5Stenelmis sandersoni5Stenelmis vittipennis5Stenochironomus4Stenonema3Strophopteryx3Strophopteryx3Strophopteryx3Stylogomphus1Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Species	Tolerance Value
Simulium venustum5Simulium vittatum7SIPHLONURIDAE7Siphloplecton2Slavina appendiculata6Somatochlora1Sperchon4Sphaerium6Sphaerium striatinum6Spinosperma ferox6catascopium6Stempellinella3Stenacron carolina2Stenacron interpunctatum7Stenelmis bicarinata5Stenelmis crenata5Stenelmis sandersoni5Stenochironomus4Stenonema3Stenonema3Stenonema3Stenohypoteryx3Stophopteryx3Stylogomphus1Stylogomphus1Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	SIMULIIDAE	6
Simulium vittatum7SIPHLONURIDAE7Siphlonurus4Siphloplecton2Slavina appendiculata6Somatochlora1Sperchon4Sphaerium6Sphaerium striatinum6Spirosperma ferox6catascopium6Stagnicola elodes6Stenacron3Stenacron carolina2Stenacron carolina2Stenelmis bicarinata5Stenelmis crenata5Stenelmis sandersoni5Stenochironomus4Stenochironomus4Strophopteryx3Strophopteryx3Stylodrilus heringianus5Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Simulium	5
SIPHLONURIDAE7Siphlonurus4Siphloplecton2Slavina appendiculata6Somatochlora1Sperchon4Sphaerium6Sphaerium striatinum6Spinosperma ferox6catascopium6Stagnicola elodes6Stenacron3Stenacron carolina2Stenacron carolina2Stenelmis bicarinata5Stenelmis crenata5Stenelmis sandersoni5Stenochironomus4Stenonema femoratum3Stenonema femoratum3Stophopteryx3Stophopteryx3Stylodrilus heringianus5Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Simulium venustum	5
Siphlonurus4Siphloplecton2Slavina appendiculata6Somatochlora1Sperchon4Sphaerium6Sphaerium striatinum6Sphaerium striatinum6Sphaerium striatinum6Spirosperma ferox6catascopium6Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenacron interpunctatum7Stenelmis bicarinata5Stenelmis crenata5Stenelmis sandersoni5Stenochironomus4Stenonema3Stenonema femoratum3Stictochironomus4Strophopteryx3Strophopteryx3Stylaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Simulium vittatum	7
Siphloplecton2Slavina appendiculata6Somatochlora1Sperchon4Sphaerium6Sphaerium striatinum6Spirosperma ferox6catascopium6Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenelmis bicarinata5Stenelmis crenata5Stenelmis sandersoni5Stenochironomus4Stenonema3Stenonema3Stictochironomus4Strophopteryx3Strophopteryx3Stylodrilus heringianus5Stylodrilus heringianus5Stylodrilus heringianus5Stylogomphus1Stylodrilus heringianus2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	SIPHLONURIDAE	7
Siphloplecton2Slavina appendiculata6Somatochlora1Sperchon4Sphaerium6Sphaerium striatinum6Spirosperma ferox6catascopium6Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenelmis bicarinata5Stenelmis crenata5Stenelmis sandersoni5Stenochironomus4Stenonema3Stenonema3Stictochironomus4Strophopteryx3Strophopteryx3Stylodrilus heringianus5Stylodrilus heringianus5Stylodrilus heringianus5Stylogomphus1Stylodrilus heringianus2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Siphlonurus	4
Slavina appendiculata6Somatochlora1Sperchon4Sphaerium6Sphaerium striatinum6Spirosperma ferox6catascopium6Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis vittipennis5Stenochironomus4Stenonema3Stophopteryx3Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		2
Sperchon4Sphaerium6Sphaerium striatinum6Spirosperma ferox6catascopium6Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenacron interpunctatum7Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis sandersoni5Stenochironomus4Stenonema3Stenonema3Stophopteryx3Strophopteryx3Stylodrilus heringianus5Stylogomphus1Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		6
Sphaerium6Sphaerium striatinum6Spirosperma ferox6catascopium6Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenacron interpunctatum7Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis sandersoni5Stenochironomus4Stenonema3Stenonema3Stophopteryx3Strophopteryx3Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Somatochlora	1
Sphaerium striatinum6Spirosperma ferox6catascopium6Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenacron interpunctatum7Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis sandersoni5Stenochironomus4Stenonema3Stenonema3Stophopteryx3Strophopteryx3Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Sperchon	4
Spirosperma ferox6Spirosperma ferox6catascopium6Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenacron interpunctatum7Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis sandersoni5Stenochironomus4Stenonema3Stenonema3Strophopteryx3Strophopteryx fasciata3Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Sphaerium	6
Spirosperma ferox6catascopium6Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenacron interpunctatum7Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis sandersoni5Stenochironomus4Stenonema3Stenonema femoratum3Stophopteryx3Strophopteryx fasciata3Stylogomphus1Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Sphaerium striatinum	6
catascopium6Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenacron interpunctatum7Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis musgravei5Stenelmis vittipennis5Stenochironomus4Stenonema3Stenonema femoratum3Strophopteryx3Strophopteryx3Styldaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		6
Stagnicola elodes6Stempellinella3Stenacron3Stenacron carolina2Stenacron interpunctatum7Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis musgravei5Stenelmis vittipennis5Stenochironomus4Stenonema3Stenonema femoratum3Stophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		6
Stempellinella3Stenacron3Stenacron carolina2Stenacron interpunctatum7Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis musgravei5Stenelmis sandersoni5Stenochironomus4Stenonema3Stenonema3Stophopteryx3Strophopteryx3Stylaria lacustris8Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		6
Stenacron3Stenacron carolina2Stenacron interpunctatum7Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis musgravei5Stenelmis sandersoni5Stenelmis vittipennis5Stenochironomus4Stenonema3Stenonema3Stophopteryx3Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	-	3
Stenacron carolina2Stenacron interpunctatum7Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis crenata5Stenelmis musgravei5Stenelmis sandersoni5Stenelmis vittipennis5Stenochironomus4Stenonema3Stenonema femoratum3Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		
Stenacron interpunctatum7Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis crenata5Stenelmis musgravei5Stenelmis sandersoni5Stenelmis vittipennis5Stenochironomus4Stenonema3Stenonema femoratum3Strophopteryx3Strophopteryx3Stylaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Stenacron carolina	
Stenelmis5Stenelmis bicarinata5Stenelmis crenata5Stenelmis musgravei5Stenelmis sandersoni5Stenelmis vittipennis5Stenochironomus4Stenonema3Stenonema femoratum3Strophopteryx3Strophopteryx3Stylaria lacustris8Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Stenacron interpunctatum	
Stenelmis bicarinata5Stenelmis crenata5Stenelmis musgravei5Stenelmis sandersoni5Stenelmis vittipennis5Stenochironomus4Stenonema3Stenonema femoratum3Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		5
Stenelmis crenata5Stenelmis musgravei5Stenelmis sandersoni5Stenelmis vittipennis5Stenochironomus4Stenonema3Stenonema femoratum3Stictochironomus4Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		
Stenelmis musgravei5Stenelmis sandersoni5Stenelmis vittipennis5Stenochironomus4Stenonema3Stenonema femoratum3Stictochironomus4Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Stenelmis crenata	
Stenelmis sandersoni5Stenelmis vittipennis5Stenochironomus4Stenonema3Stenonema femoratum3Stictochironomus4Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		
Stenelmis vittipennis5Stenochironomus4Stenonema3Stenonema femoratum3Stictochironomus4Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		
Stenochironomus4Stenonema3Stenonema femoratum3Stictochironomus4Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		
Stenonema femoratum3Stictochironomus4Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	-	
Stenonema femoratum3Stictochironomus4Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Stenonema	3
Stictochironomus4Strophopteryx3Strophopteryx fasciata3Stylaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		3
Strophopteryx fasciata3Stylaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		4
Strophopteryx fasciata3Stylaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Strophoptervx	3
Stylaria lacustris8Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	-	
Stylodrilus heringianus5Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		8
Stylogomphus1Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	-	5
Stylurus4Sublettea coffmani2Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		1
Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2		4
Sweltsa0Sympetrum10SYRPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Sublettea coffmani	2
YPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Sweltsa	
YPHIDAE10TABANIDAE6Tabanus5TAENIOPTERYGIDAE2	Sympetrum	10
Tabanus5TAENIOPTERYGIDAE2		
TAENIOPTERYGIDAE 2		
TAENIOPTERYGIDAE 2	Tabanus	5
	TAENIOPTERYGIDAE	
Taeniopteryx 2		2
Taeniopteryx burksi 2	· · ·	
Taeniopteryx nivalis 2		
Taeniopteryx parvula 2		
TALITRIDAE 8		
Tanypus 9		
Tanypus neopunctipennis 8		
	Tanytarsus	4

Species	Tolerance Value
Telopelopia okoboji	4
Thienemanniella	4
Thienemanniella similis	2
Thienemanniella xena	4
Tipula	7
Tipula abdominalis	4
TIPULIDAE	3
Tribelos	5
Trichocorixa	5
Trichocorixa calva	4
Trichocorixa kanza	4
Trichocorixa sexcincta	4
TRICORYTHIDAE	4
Tricorythodes	3
Tubifex	10
Tubifex tubifex	10
TUBIFICIDAE	10
TURBELLARIA	4
Tvetenia	5
Ulomorpha	4
UNIONIDAE	6
Valvata	8
Valvata lewisi	8
Valvata piscinalis	8
Valvata sincera	8
Valvata tricarinata	8
VALVATIDAE	8
Vejdovskyella	6
Vejdovskyella intermedia	6
VIVIPARIDAE	6
Viviparus georgianus	6
Wormaldia	2
Xenochironomus xenolabis	0
Xylotopus	2
Zavrelimyia	4

Table 7.—Tolerance values used in mIBI/HBI calculations (con't).

	BUC 15.2	BUC 14.9	BUC 13.8	BUC 11.3	BUC 9.5	BUC 9.2	BUC 8.0
mIBI Submetrics							
Total # of Taxa	3	3	3	3	3	5	3
Total Abundance	1	1	1	1	1	3	3
Number EPT Taxa	1	3	1	1	1	3	1
% Orthocladiinae & Tanytarsini	5	5	3	5	3	3	5
% Non-Insects (minus Crayfish	5	5	3	5	5	3	1
# Diptera Taxa	1	3	1	3	3	5	3
% Intolerant Taxa (Score 0-3)	3	1	1	1	1	1	1
% Tolerant Taxa (Score 8-10)	5	5	1	5	5	1	1
% Predators	3	3	3	3	5	3	3
% Shredders & Scrapers	1	1	5	1	1	5	5
% Collector/Filterers	5	3	5	5	5	5	5
% Sprawlers	1	1	1	1	1	1	1
	34	34	28	34	34	38	32
	Poor	Poor	Poor	Poor	Poor	Fair	Poor
Stand Alone Indices							
Hilsenhoff Index	4.93	4.77	6.60	5.00	4.44	6.05	6.38
	Good	Good	Fairly Poor	Good	Very Good	Fair	Fair
Shannon Index of Diversity (H')	2.55	2.77	2.59	3.00	2.39	3.07	2.47
Shannon Evenness Index (J')	0.84	0.87	0.80	0.93	0.73	0.80	0.76
% Dominance of Top 3 Taxa	0.49	0.36	0.53	0.31	0.60	0.47	0.56
% Chironomidae	0.00	0.12	0.03	0.20	0.08	0.12	0.13
QHEI Scores	66.0	64.75	59.25	39.5	51	49.3	46
	Fair	Fair	Fair	Poor	Poor	Poor	Poor

Table 8.—Scores for macroinvertebrate sites, 2018.

Table 8.—Scores for macroinvertebrate sites, 2018 (con't).

	BUC 5.9	BUC 4.0	BUC 0.9	BUC 0.2	BUC 0.0	GRE 0.1	MUN 2.2
mIBI Submetrics							
Total # of Taxa	3	3	1	3	3	5	1
Total Abundance	1	1	1	1	1	5	1
Number EPT Taxa	1	1	1	3	1	1	1
% Orthocladiinae & Tanytarsini	5	5	5	5	5	3	5
% Non-Insects (minus Crayfish	1	5	3	3	5	5	1
# Diptera Taxa	3	3	1	1	3	5	1
% Intolerant Taxa (Score 0-3)	1	1	1	3	1	1	1
% Tolerant Taxa (Score 8-10)	1	5	5	1	3	3	1
% Predators	1	3	3	1	1	3	1
% Shredders & Scrapers	5	3	1	5	5	1	5
% Collector/Filterers	5	5	5	5	3	5	5
% Sprawlers	1	1	1	1	3	1	1
	28	36	28	32	34	38	24
	Poor	Fair	Poor	Poor	Poor	Fair	Poor
Stand Alone Indices							
Hilsenhoff Index	6.85	5.65	5.63	5.57	5.24	6.96	7.64
	Fairly Poor	Fair	Fair	Fair	Good	Fairly Poor	Poor
Shannon Index of Diversity (H')	2.48	2.89	2.52	2.66	3.14	3.47	2.22
Shannon Evenness Index (J')	0.74	0.87	0.89	0.83	0.93	0.85	0.89
% Dominance of Top 3 Taxa	0.57	0.40	0.48	0.50	0.30	0.27	0.58
% Chironomidae	0.24	0.49	0.09	0.00	0.28	0.17	0.04
QHEI Scores	55.3	57.75	70.5	65.25	73	46.25	43.5
	Fair	Fair	Fair	Fair	Good	Poor	Poor

	MUN 0.1	WHI 333.4	WHI 328.1	WHI 326.9	WHI 322.2	WHI 320.1	WHI 318.8
mIBI Submetrics							
Total # of Taxa	1	5	5	5	5	3	3
Total Abundance	1	5	3	5	5	1	1
Number EPT Taxa	1	5	5	5	5	3	3
% Orthocladiinae & Tanytarsini	5	5	1	3	5	1	5
% Non-Insects (-Crayfish)	5	5	5	5	3	5	5
# Diptera Taxa	1	1	3	3	3	1	1
% Intolerant Taxa (Score 0-3)	1	5	5	5	5	3	3
% Tolerant Taxa (Score 8-10)	5	5	5	5	3	5	5
% Predators	1	1	1	1	1	1	5
% Shredders & Scrapers	1	5	1	5	3	1	1
% Collector/Filterers	3	5	1	3	3	5	5
% Sprawlers	1	1	1	1	1	1	1
	26	48	36	46	42	30	38
	Poor	Good	Fair	Good	Fair	Poor	Fair
Stand Alone Indices							
Hilsenhoff Index	5.67	4.23	4.16	4.24	4.86	4.33	3.56
	Fair	Very Good	Very Good	Very Good	Good	Very Good	Very Good
Shannon Index of Diversity (H')	1.09	3.29	3.47	3.41	3.24	3.12	2.44
Shannon Evenness Index (J')	0.79	0.84	0.88	0.84	0.79	0.91	0.77
% Dominance of Top 3 Taxa	0.90	0.34	0.22	0.31	0.34	0.29	0.54
% Chironomidae	0.00	0.00	0.10	0.07	0.04	0.03	0.00
QHEI Scores	55.8	75.25	80.75	72	66.5	66.75	69.00
	Fair	Good	Good	Good	Fair	Fair	Fair

Table 8.—Scores for macroinvertebrate sites, 2018 (con't).

Table 8.—Scores for macroinvertebrate sites, 2018 (con't).

	WHI 318.3	WHI 317.6	WHI 317.2	2 WHI 315.8	WHI 315.0	WHI 313.4	WHI 311.7
mIBI Submetrics							
Total # of Taxa	3	5	3	3	1	5	3
Total Abundance	1	5	1	1	1	3	1
Number EPT Taxa	3	3	3	3	3	5	3
% Orthocladiinae & Tanytarsini	5	3	5	5	5	5	5
% Non-Insects (-Crayfish)	5	1	5	5	5	1	3
# Diptera Taxa	1	3	1	1	1	5	3
% Intolerant Taxa (Score 0-3)	5	3	1	5	5	3	3
% Tolerant Taxa (Score 8-10)	5	1	5	5	5	3	3
% Predators	1	1	1	1	1	1	1
% Shredders & Scrapers	3	3	3	5	1	5	3
% Collector/Filterers	5	5	5	5	5	3	5
% Sprawlers	1	1	3	1	1	1	1
	38	34	36	40	34	40	34
	Fair	Poor	Fair	Fair	Poor	Fair	Poor
Stand Alone Indices							
Hilsenhoff Index	3.71	6.06	5.15	3.81	3.33	5.63	5.22
	Very Good	Fair	Good	Very Good	Excellent	Fair	Good
Shannon Index of Diversity (H')	2.90	2.91	3.13	2.65	1.74	3.23	3.22
Shannon Evenness Index (J')	0.91	0.73	0.86	0.82	0.59	0.82	0.89
% Dominance of Top 3 Taxa	0.36	0.50	0.39	0.46	0.59	0.40	0.31
% Chironomidae	0.04	0.06	0.00	0.00	0.01	0.23	0.10
QHEI Scores	68.25	66.75	61.5	75.3	60.5	65.8	58.8
	Fair	Fair	Fair	Good	Fair	Fair	Fair

	WHI 310.7	WHI 308.7	WHI 306.5	WHI 304.4	YOR 8.6	YOR 7.4	YOR 6.3
mIBI Submetrics							
Total # of Taxa	3	5	5	3	1	3	3
Total Abundance	3	5	3	3	1	1	1
Number EPT Taxa	1	5	5	3	1	3	3
% Orthocladiinae & Tanytarsini	5	3	3	5	5	5	5
% Non-Insects (-Crayfish)	3	3	1	1	5	5	3
# Diptera Taxa	3	3	1	1	1	1	1
% Intolerant Taxa (Score 0-3)	3	5	5	3	1	1	1
% Tolerant Taxa (Score 8-10)	5	5	3	5	5	3	5
% Predators	5	1	1	1	1	1	3
% Shredders & Scrapers	5	5	5	5	1	3	1
% Collector/Filterers	5	5	3	5	5	5	3
% Sprawlers	1	1	1	1	1	1	1
	42	46	36	36	28	32	30
	Fair	Good	Fair	Fair	Poor	Poor	Poor
Stand Alone Indices							
Hilsenhoff Index	4.66	4.51	4.86	5.22	6.00	6.36	5.33
	Good	Very Good	Good	Good	Fair	Fair	Good
Shannon Index of Diversity (H')	2.60	3.47	3.30	2.61	1.35	2.45	2.90
Shannon Evenness Index (J')	0.74	0.83	0.87	0.71	0.51	0.81	0.91
% Dominance of Top 3 Taxa	0.56	0.34	0.32	0.58	0.55	0.55	0.35
% Chironomidae	0.06	0.06	0.03	0.04	0.00	0.06	0.00
QHEI Scores	69.5	75.8	82.0	79.8	43.75	46.8	66.5
	Fair	Good	Good	Good	Poor	Poor	Fair

Table 8.—Scores for macroinvertebrate sites, 2018 (con't).

Table 9.—Mean scores for macroinvertebrate metrics, 2018.

Mean Scores	mIBI	Rating
WFWR Upstream of Muncie	40.4	Fair
WFWR Within Muncie	36.3	Fair
WFWR Downstream of Muncie	40.0	Fair
Buck Creek	32.7	Poor

Mean Scores	% Dom
WFWR Upstream of Muncie	0.30
WFWR Within Muncie	0.43
WFWR Downstream of Muncie	0.45
Buck Creek	0.5

Mean Scores	HBI	Rating
WFWR Upstream of Muncie	4.36	Very Good
WFWR Within Muncie	4.63	Good
WFWR Downstream of Muncie	4.81	Good
Buck Creek	5.6	Fair

Mean Scores	% Chiron.
WFWR Upstream of Muncie	0.05
WFWR Within Muncie	0.06
WFWR Downstream of Muncie	0.04
Buck Creek	0.1

Mean Scores	Η'
WFWR Upstream of Muncie	3.31
WFWR Within Muncie	2.82
WFWR Downstream of Muncie	3.00
Buck Creek	2.71

Mean Scores	J'
WFWR Upstream of Muncie	0.85
WFWR Within Muncie	0.80
WFWR Downstream of Muncie	0.79
Buck Creek	0.8

Mean Scores	QHEI	Rating	
WFWR Upstream of Muncie	72.3	Good	
WFWR Within Muncie	65.3	Fair	
WFWR Downstream of Muncie	76.8	Good	
Buck Creek	58.1	Fair	

Table 10.—Field sheet for all macroinvertebrate sampling.

Bureau of Water Quality Macroinvertebrate Sampling Field Sheet

Name of Stream Collection Date Sample ID Number of Samples Collection Notes		_Station County Method		
If riffle present score i	t 1 then rank all other habitat present			
	Natural Riffle Artificial Riffle (Rip/Rap) Slab/Bedrock w/ silt cover Cobble w/ silt cover Gravel w/ silt cover Sand w/ silt cover Mud/Silt Undercut Banks (Trees, roots, root w Riparian Vegetation (e.g. Grass) Water Willow, Root Mats Leaf Mats Logs/Woody Debris Submerged Macrophytes Filatementous Algae/Duckweed Other	vads)	w/out silt cover w/out silt cover w/out silt cover w/out silt cover	
Undercut?	No Mean depth Slight Mean width		Aesthetics Foam Discoloration	
Water Clarity	Very Max depth High water mark Clear Slight Turbid Turbid		Foam/Scum Oil Sheen Trash/Litter Nuisance Odor Sludge deposits CSOs/SSOs/Outfalls	
Incident Radiation	<u>%</u>		Impoundment Bridge	
	e there would be if the sun was directly overhead verticle incidence, canopy cover	d	Sample in lab	Date/Initials
			Macro I.D. Chironomid I.D.	

Macro taxa entered Chiron taxa entered Taxa proofed

REFERENCES

Abel, P.D. 1989. Water Pollution Biology. Ellis Horwood. Chichester, England.

- Baldigo, B.P., K. Riva-Murray, & G.E. Schuler. 2004. Effects of environmental and spatial features on mussel populations and communities in a North American river. Walkerana 14:1-32.
- Ball, B. & B. Schoenung. 1995. Recruitment of young mussels of nine commercially-valuable species in Indiana rivers. Loose-leaf publication.
- Barbour, M.T., J. Gerritsen, B.D. Snyder & J.B. Stribling. 1999. Rapid bioassessment protocols for use in streams and wadeable rivers: periphyton, benthic macroinvertebrates and fish, second edition. IPA 841-B-99-002. U.S. Environmental Protection Agency; Office of Water; Washington D.C.
- Besser, J.M, W.G. Brumbaugh, D.K. Hardesty, J.P. Hughes, & C.G. Ingersoll. 2009. Assessment of metal-contaminated sediments from the Southeast Missouri (SEMO) mining district using sediment toxicity tests with amphipods and freshwater mussels. United States Geological Society Administrative Report 08-NRDAR-02.
- Biggins, R.G., R.J. Neves & C.K. Dohner. 1995. National Strategy for the Conservation of Native Freshwater Mussels. U.S. Fish and Wildlife Service, Washington, D.C.
- Bowley, L. 2012. Bureau of Water Quality Annual Macroinvertebrate Community Report. Loose-leaf pub., n.p.
- Bowley, L. 2015. Bureau of Water Quality Annual Macroinvertebrate Community Report. Loose-leaf pub., n.p.
- Box, J.B. & J. Mossa. 1999. Sediment, land use, and freshwater mussels: prospects and problems. Journal of North American Benthological Society 18(1):99-117.
- Bringolf, R.B., W.G. Cope, C.B. Eads, P.R. Lazarno, M.C. Barnhart, & D. Shea. 2007. Acute and chronic toxicity of technical-grade pesticides to glochidia and juveniles of freshwater mussels (Unionidae). Environmental Toxicology and Chemistry 26(10):2086-2093.
- Burky, A.J. 1983. Physiological ecology of freshwater bivalves. Pages 281-387 *in* The Mollusca. Vol. 6: Ecology: ed. E.D. Russell-Hunter. Academic Press, New York, New York.
- Chutter, F.M. 1972. An empirical biotic index of the quality of water in South African streams and rivers. Water Research 6:19-30.
- Clarke, A.H. 1981. The Freshwater Molluscs of Canada. National Museums of Canada, Ottawa, Canada...
- Conrad, R.C. & S.S. Warrner. 2005. Fish Community Report, 2004. Bureau of Water Quality. Loose-leaf pub. n.p.
- Cope, W.G., R.B. Bringolf, D.B. Buchwalter, T.J. Newton, C.G. Ingersoll, N. Wing, T. Augspurger, F.J. Dwyer, M.C. Barnhart, R.J. Neves, & E. Hammer. 2008. Differential exposure, duration, & sensitivity of unionoidean bivalve life stages to environmental contaminants. Journal of the North American Benthological Society 27(2):451-462.
- Couch, K.J. 1997. An Illustrated Guide to the Unionid Mussels of Kansas. Self Published, Olathe, Kansas.
- Craddock, J.M., director. 1990. Bureau of Water Quality industrial pretreatment annual report. Loose-leaf pub, n.p.
- Dean, J., D. Edds, D. Gilette, J. Howard, S. Sherraden, & J. Tiemann. 2002. Effects of lowhead dams on freshwater mussels in the Neosho River, Kansas. Transactions of the Kansas Academy of Science
- Ellis, M.M. 1936. Erosion silt as a factor in aquatic environments. Ecology. 17:29-42.
- Epler, J.H. 2001. Identification Manual for the Larval Chironomidae (Diptera) of North and South Carolina. EPA Grant #X984170-97. North Carolina Department of Environment and Natural Resources, Division of Water Quality.

Galbraith, H.S. & C.C. Vaughn. 2011. Effects of reservoir management on abundance, condition, parasitism and reproductive traits of downstream mussels. River Research and Applications 27

Bureau of Water Quality - Habitat Summary

QHEI Score: 46.8

STREAM: YPC DATE: 7/11		STATION: SCORERS NAME:	300 W. LAB	SAMPLE#: SITE ID:	YOR - 8.6
1)SUBSTRATE (cheat TYPE Boulder/Slat Boulder [9] Cobble [8] Hardpan [4] Muck [2] Silt [2] NUMBER OF SUBST (high quality only, s) COMMENTS:	10 10		Check one, or avg. Riffle % SUBSTRATE ORIC Limestone [1] // Tills [1] Wetlands [0] Hardpan [0] Rip/Rap [0] Lacustrine [0] Shale [-1] Coal Fines [-2]	GIN SUBSTRATE QUALITY Silt Heavy [-2] Silt Moderate [-1] Silt Normal [0] Silt Free [1] EMBEDDEDNESS Extensive [-2] ✓ Moderate [-1] ✓ Normal [0]	Substrate 2.0 Max 20
2)INSTREAM COVER Undercut Banks Overhanging Veg Shallows in Slow Rootmats [1] COMMENTS:	getation [1] 🛛 🗹 Rootwa	li that occur) ▶ 70 cm [2] Oxbows, Bacl ads [1] ☑ Aquatic Macro	waters [1] Exte pphytes [1] ✓ Mod ly Debris [1] Spar	(check one, or average two) nsive > 75% [11] erate 25-75% [7] rse 5-25% [3] rly Absent < 5% [1]	Cover 15.0 Max 20
SINUOSITY [High [4] Moderate [3]	Excellent [7] No Good [5] Re ✓ Fair [3] Re	NELIZATION STABIL one [6] Hig scovered [4] V Mo	h [3] Snagging derate [2] Relocatio v [1] Canopy f Dredging	DIFICATIONS/OTHER g Impounded on Islands Removal Leveed	Channel 7.0 Max 20
4) RIPARIAN ZONE / RIPARIAN WIDTH L R Moderate 10 ✓ ✓ Narrow 5-10 ✓ Very Narrow None [0] COMMENTS:	FLÖOD L R 1 [4] - Fo 1-50 m [3] - Sh 1 m [2] - Fe	ck one per category, or two a D PLAIN QUALITY (Past 100 f rest, Swamp [3] / rub or Old Field [2] / nced Pasture [1] / nservation Tillage [1]		Image: second	Riparian] 4.8 Max 10
5) POOL/GLIDE AND MAX. DEPTH (check > 1 m [6] ✓ 0.7-1 m [4] 0.4-0.7 m [2] 0.2-0.4 m [1] < 0.2 m [POOL= COMMENTS:	 ✓ Pool w Pool w Pool w 	OGY (check one, or average idth > Riffle Width [2] idth = Riffle Width [1] idth < Riffle Width [0]	Check all that (CURRENT VE Eddies [1 Fast [1] Slow [1]	LOCITY (Pools and Riffles) I] Torrential [-1] Interstitial [-1]	Pool/Current 8.0 Max 12
Check one. RIFFLE DEPTH Best > 10 cm [2] Best 5-10 cm [1] Best < 5 cm [RIF COMMENTS: 1 6) GRADIENT (feet/	⊡ Max. < 50 c FLE=0] No Riffle [Metric = 0]	m [1] 🗌 Mod. Stable (I	ISTRATE e/boulder) [2] large gravel, sand) [1] gravel, sand) [0] 5 % Pool 2		Riffle/Run 4.0 Max 8 Gradient 6.0
			% Riffle 1	0 % Run 70	Max 10

(2):193-201.

	YPC	STATION:	Storer	SAMPLE#:	
DATE:	8/3/18	SCORERS NAME:	LAB	SITE ID: Y	OR - 8.6
400100000	- (0	
TYPE		substrate TYPE boxes) ol % Riffle % Pool %		Check one, or avg. two SUBSTRATE QUALITY	
	er/Slab [10]				
				Silt Heavy [-2]	
Bould			70 Tills [1]	Silt Moderate [-1]	Substrate
		Bedrock [5]	Wetlands [0]	Silt Normal [0]	40.0
	xan [4]	Detritus [3]	Hardpan [0]	└─ Silt Free [1]	12.0
H H Muck	[2]		Sandstone [0]	EMBEDDEDNESS	
🗀 🖳 Silt [2]		Rip/Rap [0]	Extensive [-2]	Max 20
			Lacustrine [0]	Moderate [-1]	
NUMBER OF	SUBSTRATE TYP		Shale [-1]	Normal [0]	
(high quality o	only, score > 5)	3 or less [0]	Coal Fines [-2]	None [1]	
COMMENTS:					
ZJINGTREAM	COVER (check al	TYPE (score all that occur)	Amount (check or	ie, or average two)	Cover
Undercut I	Banke [1]	Pools > 70 cm [2] Oxbows, Ba			COTCI
1.7	ing Vegetation [1]	Rootwads [1]	,		12.0
1					12.0
↓	n Slow Water [1]	Boulders [1] Logs or Woo	bdy Debris [1]		
	[1]		Nearly Absen	t < 5% [1]	Max 20
COMMENTS:					
3) CHANNEL I	MORPHOLOGY (check one per category, or average two)	Check all that occur.		
SINUOSITY	DEVELOP			ONS/OTHER	Channel
High [4]			igh [3] Snagging		
			oderate [2] Relocation		8.0
			1	1	0.0
Low [2]	└── Fair [3]		w [1] Canopy Removal		
None [1]	Poor [1	Little Recovery [1]		Bank Shaping	Max 20
COMMENTS:			One Side Channe	Modifications	
4) RIPARIAN 2	ZONE AND BANK	EROSION (check one per category, or two	and average) Right and Lef	t are facing downstream	
RIPARIAN WI		FLOOD PLAIN QUALITY (Past 100		BANK EROSION	Riparian
டக		•			•
	> 50 m [4]	L R L Eorest Swamp [3]	R Residential Park New Field [1]	L R ✓ ✓ None/Little [3]	4.8
Wide	> 50 m [4] rate 10-50 m [3]	Forest, Swamp [3]	Residential, Park, New Field [1]		4.8
Wide	rate 10-50 m [3]	Forest, Swamp [3] Shrub or Old Field [2]	Urban, Industrial [0]	Moderate [2]	
Wide	rate 10-50 m [3] w 5-10 m [2]	Forest, Swamp [3] Shrub or Old Field [2] Fenced Pasture [1]	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0]		4.8 Max 10
Wide Model Model Virght	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1]	Forest, Swamp [3] Shrub or Old Field [2]	Urban, Industrial [0]	Moderate [2]	
Wide Modes Modes V Narrov Very I None	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1]	Forest, Swamp [3] Shrub or Old Field [2] Fenced Pasture [1]	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0]	Moderate [2]	
Wide Model Model Virght	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1]	Forest, Swamp [3] Shrub or Old Field [2] Fenced Pasture [1]	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0]	Moderate [2]	
Wide Model Model Very I Very I COMMENTS:	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1]	Forest, Swamp [3] Shrub or Old Field [2] Fenced Pasture [1] Conservation Tillage [1]	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0]	Moderate [2]	
Wide Model Model Very I Very I COMMENTS:	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE/	Forest, Swamp [3] Shrub or Old Field [2] Fenced Pasture [1] Conservation Tillage [1]	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur	Mone/Liftle [3]	
Wide Wide Model Very I Very I ONONE COMMENTS:	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE/	Forest, Swamp [3] Forest, Swamp [3] Fonced Pasture [1] Conservation Tillage [1] RUN QUALITY MORPHOLOGY (check one, or average	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur two) CURRENT VELOCITY	Pools and Riffles)	Max 10
Wide Wide Model Very I Very I ONONE COMMENTS:	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE/ (check one)	Forest, Swamp [3] Shrub or Old Field [2] Fenced Pasture [1] Conservation Tillage [1] MORPHOLOGY (check one, or average Pool width > Riffle Width [2]	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur two) CURRENT VELOCITY Eddies [1]	Pools and Riffles)	Max 10 Pool/Current
↓ Wide ↓ Modes ↓ Narros ↓ Very f ↓ None COMMENTS: S) POOL/GLIC MAX. DEPTH > 1 m [6] ↓ 0.7-1 m [4]	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE/ (check one)]	Forest, Swamp [3] Forest, Swamp [3] Shrub or Old Field [2] Fenced Pasture [1] Conservation Tillage [1] MORPHOLOGY (check one, or average Pool width > Riffle Width [2] Pool width = Riffle Width [1]		Peools and Riffles)	Max 10
↓ Wide ↓ Modes ↓ Narroo ↓ Very f ∧ None COMMENTS: S) POOL/GLIC MAX. DEPTH > 1 m [6] 0.7-1 m [4] 0.4-0.7 m	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE / (check one)] [2]	Forest, Swamp [3] Shrub or Old Field [2] Fenced Pasture [1] Conservation Tillage [1] MORPHOLOGY (check one, or average Pool width > Riffle Width [2]		Peools and Riffles) Torrential [-1] Interstitial [-2]	Max 10 Pool/Current 0.0
↓ Wide ↓ Model ✓ Narro ✓ Very I ✓ None COMMENTS: S) 5) POOL/GLIC MAX. DEPTH > 1 m [6] ○.7-1 m [4] 0.4-0.7 m ○.2-0.4 m □	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE / (check one)] [2] [1]	Forest, Swamp [3] Forest, Swamp [3] Shrub or Old Field [2] Fenced Pasture [1] Conservation Tillage [1] MORPHOLOGY (check one, or average Pool width > Riffle Width [2] Pool width = Riffle Width [1]		Peools and Riffles)	Max 10 Pool/Current
↓ Wide ↓ Model ✓ Narro ✓ None COMMENTS: S) POOL/GLID MAX. DEPTH > 1 m [6] 0.7-1 m [4] 0.4-0.7 m Ø.2-0.4 m < 0.2 m [P	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE / (check one)] [2] [1]	Forest, Swamp [3] Forest, Swamp [3] Shrub or Old Field [2] Fenced Pasture [1] Conservation Tillage [1] MORPHOLOGY (check one, or average Pool width > Riffle Width [2] Pool width = Riffle Width [1]		Peools and Riffles) Torrential [-1] Interstitial [-2]	Max 10 Pool/Current 0.0
↓ Wide ↓ Model ✓ Narro ✓ Very I ✓ None COMMENTS: S) 5) POOL/GLIC MAX. DEPTH > 1 m [6] ○.7-1 m [4] 0.4-0.7 m ○.2-0.4 m □	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE / (check one)] [2] [1]	Forest, Swamp [3] Shrub or Old Field [2] Fenced Pasture [1] Conservation Tillage [1] KUN QUALITY MORPHOLOGY (check one, or average Pool width > Riffle Width [2] Pool width < Riffle Width [1]		Peools and Riffles) Torrential [-1] Interstitial [-2]	Max 10 Pool/Current 0.0
↓ Wide ↓ Model ✓ Narro ✓ Very I ✓ None COMMENTS: MAX. DEPTH → 1 m [6] 0.7-1 m [4] 0.4-0.7 m ✓ 0.2-0.4 m ✓ < 0.2 m [P	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE / (check one)] [2] [1] 'OOL=0]	Image: Second system [3] Image: Second system <td< td=""><td>Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur two CURRENT VELOCITY Eddies [1] Fast [1] Moderate [1] Slow [1]</td><td>Pools and Riffles) Torrential [-1] Interstitial [-1] Intermittent [-2] Very Fast [1]</td><td>Max 10 Pool/Current 0.0 Max 12</td></td<>	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur two CURRENT VELOCITY Eddies [1] Fast [1] Moderate [1] Slow [1]	Pools and Riffles) Torrential [-1] Interstitial [-1] Intermittent [-2] Very Fast [1]	Max 10 Pool/Current 0.0 Max 12
↓ Wide ↓ Model ✓ Narro ✓ Very I ✓ None COMMENTS: None 5) POOL/GLIC MAX. DEPTH → 1 m [6] 0.7-1 m [4] 0.4-0.7 m ○ 0.2-0.4 m ✓ < 0.2 m [P	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE / (check one)] [2] [1] 'OOL=0] H	Forest, Swamp [3] Forest, Swamp [3] Shub or Old Field [2] Fenced Pasture [1] Conservation Tillage [1] WORPHOLOGY (check one, or average Pool width > Riffle Width [2] Pool width > Riffle Width [1] Pool width < Riffle Width [0]	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur two CURRENT VELOCITY Eddies [1] Fast [1] Moderate [1] Slow [1] G. two Check or BSTRATE REFLE/	Pools and Riffles) Torrential [-1] Interstitial [-1] Intermittent [-2] Very Fast [1] re, or avg. two RUN EMBEDDED.	Max 10 Pool/Current 0.0
↓ Wide ↓ Model ✓ Narro ✓ Very I ✓ None COMMENTS: None 5) POOL/GLIC MAX. DEPTH → 1 m [6] 0.7-1 m [4] 0.4-0.7 m ✓ 0.2-0.4 m ✓ 0.2 m [P COMMENTS: Check one. RIFFLE DEPT Best > 10	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE / (check one)] [2] [1] 'OOL=0] H cm [2]	Forest, Swamp [3] Forest, Swamp [3] Shub or Old Field [2] Fenced Pasture [1] Conservation Tillage [1] MORPHOLOGY (check one, or average Pool width > Riffle Width [2] Pool width > Riffle Width [1] Pool width < Riffle Width [0]	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur two) CLIRRENT VELOCITY Eddies [1] Fast [1] Moderate [1] Slow [1] G. two Check or BSTRATE RIFFLE/I le/boulder) [2] Non	Peools and Riffles) Torrential [-1] Interstitial [-1] Interstitial [-1] Very Fast [1] e, or avg. two RUN EMBEDDED. ≥ [2]	Max 10 Pool/Current 0.0 Max 12 Riffle/Run
↓ Wide ↓ Model ✓ Narro ✓ Very I ✓ None COMMENTS: None 5) POOL/GLIC MAX. DEPTH → 1 m [6] 0.7-1 m [4] 0.4-0.7 m ○ 0.2-0.4 m ✓ 0.2 m [P COMMENTS: Check one. RIFFLE DEPT Best > 10 ✓ Best 5-10	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE / (check one)] [2] [1] 'OOL=0] H cm [2] cm [1]	Image: Check one. Check one. Check one. or awarage Image: Check one. <t< td=""><td>Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur (URRENT VELOCITY Eddies [1] Fast [1] Moderate [1] Slow [1] G. two Check or BSTRATE RIFFLE/I le/boulder) [2] Non (large gravel, sand) [1] Low</td><td>(Peools and Riffles) Torrential [-1] Interstitial [-1] Interstitial [-1] Very Fast [1] Re, or avg. two RUN EMBEDDED. 2[2] [1]</td><td>Max 10 Pool/Current 0.0 Max 12</td></t<>	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur (URRENT VELOCITY Eddies [1] Fast [1] Moderate [1] Slow [1] G. two Check or BSTRATE RIFFLE/I le/boulder) [2] Non (large gravel, sand) [1] Low	(Peools and Riffles) Torrential [-1] Interstitial [-1] Interstitial [-1] Very Fast [1] Re, or avg. two RUN EMBEDDED. 2[2] [1]	Max 10 Pool/Current 0.0 Max 12
↓ Wide ↓ Model ✓ Narro ✓ Very I ✓ None COMMENTS: None 5) POOL/GLIC MAX. DEPTH → 1 m [6] 0.7-1 m [4] 0.4-0.7 m ○ 0.2-0.4 m ✓ 0.2 m [P COMMENTS: Check one. RIFFLE DEPT Best > 10 ✓ Best 5-10	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE / (check one)] [2] [1] 'OOL=0] H cm [2]	Image: Check one. Check one. Check one. or awarage Image: Check one. <t< td=""><td>Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur CURRENT VELOCITY Eddies [1] Fast [1] Moderate [1] Slow [1] G. two Check or BSTRATE RIFFLE/ le/boulder) [2] Non (large gravel, sand) [1] Low e gravel, sand) [0] Moderate [1] Low</td><td>Poole/Liftle [3] ✓ Moderate [2] Severe [1] (Peools and Riffles) Torrential [-1] Interstitial [-1] Interstitial [-1] Very Fast [1] e, or avg. two RUN EMBEDDED. ≥ [2] [1] erate [0]</td><td>Max 10 Pool/Current 0.0 Max 12 Riffle/Run 3.0</td></t<>	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur CURRENT VELOCITY Eddies [1] Fast [1] Moderate [1] Slow [1] G. two Check or BSTRATE RIFFLE/ le/boulder) [2] Non (large gravel, sand) [1] Low e gravel, sand) [0] Moderate [1] Low	Poole/Liftle [3] ✓ Moderate [2] Severe [1] (Peools and Riffles) Torrential [-1] Interstitial [-1] Interstitial [-1] Very Fast [1] e, or avg. two RUN EMBEDDED. ≥ [2] [1] erate [0]	Max 10 Pool/Current 0.0 Max 12 Riffle/Run 3.0
↓ Wide ↓ Model ✓ Narro ✓ Very I ✓ None COMMENTS: None 5) POOL/GLIC MAX. DEPTH → 1 m [6] 0.7-1 m [4] 0.4-0.7 m ○ 0.2-0.4 m ✓ 0.2 m [P COMMENTS: Check one. RIFFLE DEPT Best > 10 ✓ Best 5-10	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE / (check one)] [2] [1] 'OOL=0] H cm [2] cm [1]	Image: Second system Forest, Swamp [3] Image: Shrub or Old Field [2] Image: Shrub or Old Field Field Field [2]	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur CURRENT VELOCITY Eddies [1] Fast [1] Moderate [1] Slow [1] G. two Check or BSTRATE RIFFLE/ le/boulder) [2] Non (large gravel, sand) [1] Low e gravel, sand) [0] Moderate [1] Low	(Peools and Riffles) Torrential [-1] Interstitial [-1] Interstitial [-1] Very Fast [1] Re, or avg. two RUN EMBEDDED. 2[2] [1]	Max 10 Pool/Current 0.0 Max 12 Riffle/Run
↓ ↓ Wide ↓ ↓ Mode: ↓ ↓ Narro ↓ ↓ None COMMENTS: None ↓ ↓ 1m [6] ↓ ↓ 0.2-0.4 m ↓ <0.2 m [P	rate 10-50 m [3] w 5-10 m [2] Narrow < 5 m [1] [0] DE AND RIFFLE / (check one)] [2] [1] 'OOL=0] H cm [2] cm [1] m [RIFFLE=0]	Image: Second system Forest, Swamp [3] Image: Shrub or Old Field [2] Image: Shrub or Old Field Field Field [2]	Residential, Park, New Field [1] Urban, Industrial [0] Open Pasture, Row Crop [0] Mining, Construction [0] Check all that occur CURRENT VELOCITY Eddies [1] Fast [1] Moderate [1] Slow [1] G. two Check or BSTRATE RIFFLE/ le/boulder) [2] Non (large gravel, sand) [1] Low e gravel, sand) [0] Moderate [1] Low	Poole/Liftle [3] ✓ Moderate [2] Severe [1] (Peools and Riffles) Torrential [-1] Interstitial [-1] Interstitial [-1] Very Fast [1] e, or avg. two RUN EMBEDDED. ≥ [2] [1] erate [0]	Max 10 Pool/Current 0.0 Max 12 Riffle/Run 3.0

Gatenby, C.M., P.A. Morrison, R.J. Neves & B.C. Parker. 1998. A protocol for the salvage and

quarantine of Unionid mussels from zebra mussel-infested waters. Proceedings of the Conservation,

Bureau of Water Quality - Habitat Summary

QHEI Score: 66.5

STREAM: YPC	10	STATION:	400 W.	SAMPLE#:	VOD CO
DATE: <u>7/11/</u>	8	SCORERS NAME:	LAB	SITE ID:	YOR - 6.9
1)SUBSTRATE (check TYPE Boulder/Slab [Boulder [9] Cobble [8] Hardpan [4] Muck [2] Silt [2] NUMBER OF SUBSTR (high quality only, see COMMENTS:	10 30 ATE TYPES	Pool%	Check one, or ave Riffle % SUBSTRATE OR 60 Limestone [1 30 / Tills [1] Wetlands [0] Hardpan [0] Sandstone [[Rip/Rap [0] Lacustrine [0 Shale [-1] Coal Fines [-	SUBSTRATE QUALITY Sitt Heavy [-2] Sitt Moderate [-1] Sitt Normal [0] Sitt Free [1] BEDDEDNESS Extensive [-2] Moderate [-1] Normal [0]	Substrate 16.5 Max 20
2)INSTREAM COVER Undercut Banks [1] Overhanging Vege Shallows in Slow V Rootmats [1] COMMENTS:	tation [1] 🛛 🗌 Rootw	li that occur) > 70 cm [2] Oxbows, Bac ads [1] ✓ Aquatic Mac	kwaters [1] Ex ophytes [1] \checkmark Mc dy Debris [1] Sp	it (check one, or average two) tensive > 75% [11] oderate 25-75% [7] varse 5-25% [3] varly Absent < 5% [1]	Cover 15.0 Max 20
	Excellent [7] ✓ No Good [5] Re Fair [3] Re	NELIZATION STABI one [6] Hi ecovered [4] V M	gh [3] Snaggi oderate [2] Reloca w [1] Canopy Dredgii	DDIFICATIONS/OTHER ing Impounded tion Islands y Removal Leveed	Channel 13.0 Max 20
COMMENTS.					
4) RIPARIAN ZONE AI RIPARIAN WIDTH L R ✓ Moderate 10-5 ✓ Narrow 5-10 m Very Narrow < None [0] COMMENTS:	FLÖÖD L R 4] Fc 10 m [3] Si 1[2] Fe	eck one per category, or two a D PLAIN QUALITY (Past 100 prest, Swamp [3] Inub or Old Field [2] Inced Pasture [1] Inservation Tillage [1]		✓ ✓ Moderate [2] Crop [0] ✓ Severe [1]	
5) POOL/GLIDE AND MAX. DEPTH (check o > 1 m [6] ✓ 0.7-1 m [4] 0.4-0.7 m [2] 0.2-0.4 m [1] < 0.2 m [POOL=0] COMMENTS:	 ✓ Pool w Pool w Pool w 	.OGY (check one, or average idth > Riffle Width [2] idth = Riffle Width [1] idth < Riffle Width [0]	Eddies Fast [1 ✓ Modera ✓ Slow [1	/ELOCITY (Pools and Riffles) [1] Torrential [-1]] Interstitial [-1] ate [1] Intermittent [-2]] Very Fast [1]	Pool/Current 8.0 Max 12
Check one. RIFFLE DEPTH ☐ Best > 10 cm [2] ✓ Best 5-10 cm [1] ☐ Best < 5 cm [RIFF] COMMENTS: ☐ No	Check one. RUN DEPTH ☐ Max. > 50 d ☑ Max. < 50 d LE=0] 9 Riffle [Metric = 0]	m [1] 🗹 Mod. Stable		Check one, or avg. two RIFFLE/RUN EMBEDDED. None [2] Low [1] Moderate [0] Extensive [-1]	Riffle/Run 3.0 Max 8 Gradient
6) GRADIENT (feet/mi	le) <u>6.9</u> DRAIN	IAGE AREA (sq. mile) <u>9.</u>	0 % Pool % Riffle	30 % Glide 5 % Run 65	6.0 Max 10

Captive Care, and Propagation of Freshwater Mussel Symposium. Ohio Biological Survey, Columbus, Ohio.

- Gerritsen, J., Carlson, R.E., Dycus, D.L., Faulkner, C., Gibson, G.R., Harcum, J., & Markowitz, S.A. 1998. Lake and Reservoir Bioassessment and Biocriteria. Technical Guidance Document. US environmental Protection Agency. EPA 841-B-98-007. 10 Chapters, Appendices A-G. (http://www.epa.gov/owow/monitoring/tech/lakes.html)
- Gooding, M.P., T.J. Newton, M.R. Bartsch, & K.C. Hornbuckle. 2006. Toxicity of synthetic musks to early life stages of the freshwater mussel *Lampsilis cardium*. Archives of Environmental Contamination and Toxicology 51(4):549-558.
- Grabarkiewicz, J. & W. Davis. 2008. An Introduction to Freshwater Mussels as Biological Indicators EPA-260-R-08-015. U.S. Environmental Protection Agency, Office of Environmental Information, Washington, DC.
- Haag, W.R. 2009. Past and future patterns of freshwater mussel extinctions in North America during
- the Holocene. Pages 107-128 in Holcene Extinctions, Oxford University Press, New York.
- Hallmann, C.A., Sorg, M., Jongejans, E., Siepel, H., Hofland, N., Schwan, H., Stenmans, W., Muller, A., Sumser, H., Horren, T., Goulsen, F., and H. de Kroon. 2017. More than 75 percent decline over 27 years in total flying insect biomass in protected areas. PLoS One 12, e0185809.
- Hellawell, J. 1986. Biological Indicators of Freshwater Pollution and Environmental Management. Elsevier Applied Science Publications, Elsevier, London.
- Hickling, R., Roy David, B., Hill Jane, K., and D. Chris Thomas. A northward shift of range margins in British Odonata. Glob. Chang. Biol. 11:502-506.
- Hilsenhoff, W.L. 1987. An improved biotic index of organic stream pollution. The Great Lakes Entomologist 20:31-39.
- Hoggat, R.E. 1975. Drainage Areas of Indiana Streams. Department of Interior. Geological Survey. Water Resources Division.
- Houghton, D.C. and R.W. Holzenthal. 2010. Historical and contemporary biological diversity of Minnesota caddisflies: a case study of landscape-level species loss and trophic composition shift.
- J.N. Am. Benthol. Society. 29:480-495.
- Indiana Department of Environmental Management. 2010. Multi-habitat (MHAB) Macroinvertebrate Collection Procedure. S-001-OWQ-W-BS_10-T-R0. Technical Standing Operating Procedure. Office of Water Quality, Indianapolis, IN.
- International Council for Local Environmental Initiatives, Case Study #19 (ICLEI Case Study #19).
 1994. Local Water Pollution Control, Industrial Pretreatment & Biological Indicators.
 Irvine, J.R. 1985. Effects of successive flow perturbations on stream invertebrates. Canadian Journal of Fisheries and Aquatic Sciences. 42:1922–1927.
- Jenderedjian, K., Hakobyan, S. and M.A. Stapanian. 2012. Trends in benthic macroinvertebrate community biomass and energy budgets in Lake Sevan, 1928-2004. Environ. Monit. Assess. 184:6647-6671.
- Kadoya, T., Suda, S.-i., and I. Washitani. 2009. Dragonfly crisis in Japan: a likely consequence of recent agricultural habitat degradations. Biol. Conserv. 142:1899-1905.
- Kaesler, R.L., Herricks, E.E., & J.S Crossman. 1978. Use of Indices of Diversity and Hierarchical Diversity in Stream Surveys in Biological Data in Water Pollution Assessment; Quantitative and Statistical Analyses, ASTM STP 652. (K.L. J. Cairns, Jr. & R.J. Livingston, eds.), American Society for Testing and Materials
- Kalkman, V.j., Boudot, J.-P., Bernard, R. Conze, K.-J.r., Knijf, G.D., Dyatlova, E., Ferreira, S.n., Jovic, M., Ott, J.r. Rivervato, E., and G.r. Sahlen. 2010. European Red List of Dragonflies. Publications

Office of the European Union, Luxembourg.

- Karatayev, A.Y., Burlakova, L.E., Padilla, D.K., Mastistsky, S.E., and S. Olenin. 2009. Invaders are not a random selection of species. Biol. Invasions. 11:2009-2019.
- Karr, J.R., & D.R. Dudley. 1981. Ecological perspective on water quality goals. Environmental Management 5:55-68.
- Karr, J.R. 1981. Assessment of biotic integrity using fish communities. Fisheries 6(6):21-27.
- Karr, J.R. 1991. Biological integrity: a long-neglected aspect of water resource management. Ecological Applications 1 (1):66-84.
- Keller, A.E., T. Augspurger. 2005. Toxicity of fluoride to the endangered Unionid mussel, *Alasmidonta ravenellana*, and surrogate species. Bulletin of EnvironmentalContamination and Toxicology 74:242-249.
- Klemm, J.D., P. L. Lewis, F. Fulk, & J.M. Lazorchak. 1990. Macroinvertebrate Field and Laboratory Methods for Evaluating the Biological Integrity of Surface Waters. US EPA Publication# EPA/600/4-90/030.
- Lenat, D.R., L.A. Smock, & D.L. Penrose. 1980. Use of benthic macroinvertebrates as indicators of environmental quality. Pages 97-112 *in* Biological Monitoring for Environmental Effects. (D.L. Worf & D.C. Heath, eds.). Lexington, Massachusetts.
- Lenat, D.R. & M.T. Barbour. 1993. Using benthic Macroinvertebrate community structure for rapid, cost-effective, water quality monitoring: rapid bioassessment. Pages 187-215 *in* Biological Monitoring of Aquatic Systems. (S.L. Loeb & A. Spacie, eds.). Lewis Publishers. Boca Raton, Florida.
- Lessard, J.L. & D.B. Hayes. 2003. Effects of elevated water temperature on fish and macroinvertebrate communities below small dams. River Research and Applications 19(7):721-732.
- Lister, B.C., and A. Garcia. 2018. Climate-driven declines in arthropod abundance restructure a rainforest food web. Proc. Natl. Acad. Sci.155:44.
- Ludwig, J.A. & J.F. Reynolds. 1988. Statistical Ecology: A Primer on Methods and Computing. John Wiley & Sons, New York, New York.
- Lydeard, C., R.H. Cowie, W.F. Ponder, A.E. Bogan, P. Bouchet, S.A. Clark, K.S. Cummings, T.J. Frest, O. Gargominy, D.G. Herbert, R. Hershler, K.E. Perez, B. Roth, M. Seddon, E.E. Strong, & F.G. Thompson. 2004. The global decline of nonmarine mollusks. BioScience 54(4):321-330.
- Maloney, K.O., H.R. Dodd, S.E. Butler, & D.H. Wahl. 2008. Changes in macroinvertebrate and fish assemblages in a medium-sized river following a breach of a low-head dam. Freshwater Biology 53:1055-1063.
- March, F.A., Dwyer, F.J., Augspurger, A., Ingersoll, C.G., Wang, N., & C.A. Mebane. 2007. An evaluation of freshwater mussel toxicity data in the derivation of water quality guidance and standards for copper. Environmental Toxicology and Chemistry 26(10):2066-2074.
- Mason, W.T. 1998. Watershed Assessments with Chironomidae: Diptera. Ecology Support, Inc. Gainesville, Florida.
- McKinney, M.L. 2006. Urbanization as a major cause of biotic homogenization. Biol. Conserv. 127:2247-260.
- Mummert, A.K., R.J. Neves, T.J. Newcomb, & D.S. Cherry. 2003. Sensitivity of juvenile freshwater mussels to total un-ionized ammonia. Environmental Toxicology and Chemistry 22(11):2545-2553.
- Neves, R.J., A.E. Bogan, J.D Williams, S.A. Ahlstedt, & P.W. Hartfield. 1997. Status of aquatic mollusks in the southeastern United States: a downward spiral of diversity. Aquatic fauna in peril: the Southeastern perspective. Special Publication 1:44-86.
- NRCS. 2007. Native Freshwater Mussels. Fish and Wildlife Habitat Management Leaflet.

- Ohio Environmental Protection Agency (OEPA). 2006. Methods for Assessing Habitat in Flowing Waters: Using the Qualitative Habitat Evaluation Index (QHEI). Ohio EPA Technical Bulletin EAS/2006-06-1.
- Parmalee, P.W. & A.E. Bogan. 1998. The Freshwater Mussels of Tennessee. The University of Tennessee Press, Knoxville, Tennessee.
- Payne, B.S., A.C. Miller & L.R. Shaffer. 1999. Physiological resilience of freshwater mussels to turbulence and suspended solids. Journal of Freshwater Ecology 14(2).
- Peckarsky, B.L., P.R. Fraissinet, M.A. Penton & D.J. Conklin, Jr. 1990. Freshwater Macroinvertebrates of Northeastern North America. Cornell University Press, New York, New York.
- Pielou, E.C. 1966.. The measurement of diversity in different types of biological collections. Journal of Theoretical Biology 13:131-144.
- Plafkin, J.L., Barbour, M.T., Porter, K.D., Gross, S.K., & R.M. Hughes. 1989. Rapid Bioassessment Protocols for use in Streams and Rivers: Benthic Macroinvertebrates and Fish. U.S. Environmental Protection Agency. EPA 440/4 - 89/001. 8 chapters, Appendices A-D.
- Poff, N.L., J.D. Olden, D.M. Merritt, & D.M. Pepin. 2007. Homogenization of regional river dynamics by dams and global biodiversity implications. Proceedings of the National Academy of Sciences of the United States of America 104:5732-5737.
- Poole, K.E., & J.A. Downing. 2004. Relationship of declining mussel biodiversity to stream-reach and watershed characteristics in an agricultural landscape. Journal of the North American Benthological Society 23(1):114-125.
- Rankin, E.T. 1989. The Qualitative Habitat Evaluation Index (QHEI): Rationale, Methods, and Application. Ohio Environmental Protection Agency, Ecological Assessment Section, Division of Water Quality and Assessments, Columbus, Ohio.
- Régnier, C.B. Fontaine & P. Bouchet. 2009. Not knowing, not recording, not listing: numerous unnoticed mollusk extinctions. Conservation Biology 23(5):1214-1221.
- Ricciardi, A., & J.B. Rasmussen. 1999. Extinction rates of North American freshwater fauna. Conservation Biology 13:1220-1222.
- Roy, A.H., A.D. Rosemond, M.J. Paul, D.S. Leigh, & J.B. Wallace. 2003. Stream macroinvertebrate response to catchment urbanization. Freshwater Biology 48:329-346.
- Sanchez-Bayo, F., and K.A.G. Wyckhuys. 2019. Worldwide decline of the entomofauna: A review of its drivers. Biological Conservation. 232:8-27.
- Santucci, V J., S.R. Gephard, & S.M. Pescitelli. 2005. Effects of multiple low-head dams on fish, macroinvertebrates, habitat, and water quality in the Fox River, Illinois. North American Journal of Fisheries Management 25:975-992.
- Shortall, C.r., Moore, A., Smith, E., Hall, M.J., Woiwod, I.P., and R. Harrington. 2009. Long-term changes in the abundance of flying insects. Insect Conserv. Divers. 2:251-260.
- Smith, D.R., R.F. Villella, D.P. Lemarie, & S. von Oettingen. 1999. How much excavation is needed to monitor freshwater mussels? Proceedings of the First Freshwater Conservation Society Symposium. Ohio Biological Survey, Columbus, Ohio.
- Strayer, D.L. 1999a. Freshwater mollusks and water quality. Journal of North American Benthological Society 18(1):1.
- Strayer, D.L., & D.R. Smith. 2003. A Guide to Sampling Freshwater Mussel Populations. American Fisheries Society, Monograph 8, Bethesda, Maryland.
- Strayer, D.L., J.A. Downing, W.R. Haag, T.L. King, J.B Layzer, T.J. Newton, & S.J. Nichols. 2004. Changing perspectives on pearly mussels, North America's most imperiled animals. BioScience 54(5):429-439.

- Strayer, D.L. 2008. Freshwater Mussel Ecology: A Multifactor Approach to Distribution and Abundance. University of California Press, Berkeley and Los Angeles, California.
- Tesmer, M.G. & D.R. Wefring. 1979. Annual macroinvertebrate sampling- a low cost tool for ecological assessment of effluent impact. Pages 264-279 in Ecological Assessments of Effluent Impacts on Communities of Indigenous Aquatic Organisms. (J. M. Bates & C. I. Weber, eds.). American Society for Testing and Materials, Philadelphia, Pennsylvania.
- Thomas, J.A., Telfer, M.G., Roy, D.B., Preseton, C.D., Greenwood, J.J.D., Asher, J., Fox, R., Clarke, R.T., and J.H. Lawton. 2004. Comparative losses of British butterflies, birds, and plants and the global extinction crisis. Science. 303:1879-1881.
- Tiemann, J.S., G.P. Gillette, M.L. Wildhaber, & D.R. Edds. 2004. Effects of lowhead dams on riffledwelling fishes and macroinvertebrates in a midwestern river. Transactions of the American Fisheries Society 133:705-717.
- Tiemann, J.S., H.R. Dodd, N. Owens, & D.H. Wahl. 2007. Effects of lowhead dams on unionids in the Fox River, Illinois. Northeastern Naturalist 14(1):125-138.
- Tiemann, J.S., S.A. Douglass, A P. Stodola, & K S. Cummings. 2016. Effects of lowhead dams on freshwater mussels in the Vermilion River Basin, Illinois with comments on a natural dam removal. Transactions of the Illinois State Academy of Science 109:1-7.
- Valenti, T.W., D.S. Cherry, R.J. Neves, & J. Schmerfeld. 2005. Acute and chronic toxicity of mercury to early life stages of the Rainbow Mussel, *Villosa iris* (Bivalve: Unionidae). Environmental Toxicology and Chemistry 24(5):1242-1246.
- Vandermeer, J. 1981. Elementary Mathematical Ecology. John Wiley & Sons, USA.
- Vannote, R.L. & B.W. Sweeney. 1980. Geographic analysis of thermal equilibria: A conceptual model for evaluating the effect of natural and modified thermal regimes on aquatic insect communities. The American Naturalist. 115:667-695.
- Vaughn, C.C. & C.M. Taylor. 1999. Impoundments and the decline of freshwater mussels: a case study of an extinction gradient. Conservation Biology 13(4):912-920.
- Wang, N., C.G. Ingersoll, D.K. Hardesty, C.D. Ivey, J.L. Kunz, T.W. May, F.J. Dwyer, A.D. Roverts, T. Augspurger, C.M. Kane, R.J. Nevers & M.C. Barnhart. 2007. Acute toxicity of copper, ammonia, and chlorine to glochidia and juveniles of freshwater mussels. Environmental Toxicology and Chemistry. 26(10):2036-2047.
- Ward, J.V. 1976. Effects of thermal constancy and seasonal temperature displacement on community structure of stream macroinvertebrates. Thermal Ecology II: proceedings of a symposium held at Augusta, Georgia, April 2-5. 1975. pp. 302-307.
- Watters, G.T. 1995. A Guide to the Freshwater Mussels of Ohio. Third Edition. Ohio Division of Wildlife, Columbus, Ohio.
- Watters, G.T. 1996. Small dams as barriers to freshwater mussels (Bivalvia, Unionoida) and their hosts. Biological Conservation 75(1):79-85.
- Watters, G.T. 1999. Freshwater mussels and water quality: A review of the effects of hydrologic and instream habitat alterations. Proceedings of the First Freshwater Mollusk Conservation Society Symposium. Ohio Biological Survey. Columbus, Ohio.
- Watters, G.T. 2000. Freshwater mussels and water quality: A review of the effects of hydrologic and instream habitat alterations. Proceedings of the First Freshwater Mollusk Conservation Society Symposium. Ohio Biological Survey, Columbus, Ohio.
- Wilhm, J. 1967. Comparison of some diversity indices applied to populations of benthic macroinvertebrates in a stream receiving organic wastes. Journal of the Water Pollution Control Federation 39:1674-1683.

- Zahradkova, S., Soldan, T., Bojkova, J., Helesic, J., Janovska, H., and P. Sroka. 2009. Distribution and biology of mayflies (Ephemeroptera) of the Czech Republic: present status and perspectives. Aquat. Insects. 31:629-652.
- Zedkova, B., Radkova, B., Bojkova, J., Soldan, T., and S. Zahradkova. 2015. Mayflies (Ephemeroptera) as indicators of environmental changes in the past five decades: a case study from the Morava and Odra River Basins (Czech Republic). Aquat. Conserv. 25:622-638.